

Design, synthesis and Structure-Activity Relationship of novel Phenolic based Pyrimidine hybrids from Cashew Nut Shell Liquid (CNSL) components as potential antitumor agents

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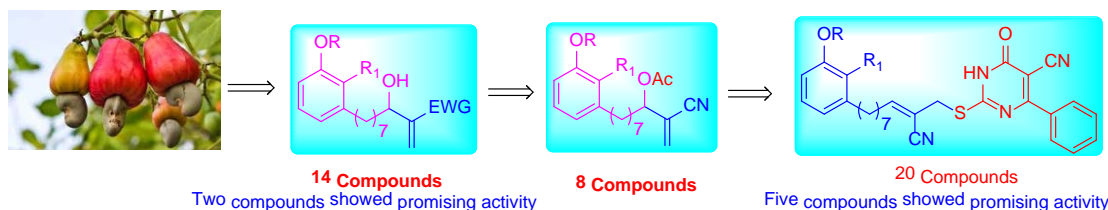
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ABSTRACT



Herein we describe a simple method for the synthesis of Baylis-Hillman adducts and their acetates by utilizing inexpensively available cashew nut shell liquid (CNSL) natural resource. Furthermore, by using a molecular hybridization approach, a series of novel pyrimidine scaffolds (**15a-15t**) were synthesized by utilizing Baylis-Hillman acetate derivatives from cashew nut shell liquid (CNSL). All the newly synthesized compounds were screened for their *in vitro* antitumor activity. Baylis-Hillman compounds **5b** and **5h** showed promising anticancer activity against MCF-7. Among pyrimidine derivatives, compounds **15i**, **15j** and **15l** showed promising activity against HEP-G2, whereas compounds **15j**, **15k**, **15l** and **15m** showed promising activity against MCF-7. In addition, compound **15m** showed significant activity against K562 when compared with the standard. The structure-activity relationship (SAR) analysis suggests that the length of the carbon chain of phenyl ring played an important role in the potency of activity.

Keywords: Cashew nut shell liquid, Baylis-Hillman adducts, pyrimidine derivatives, Anti-cancer activity

INTRODUCTION

Investigation of new chemical entities for the treatment of cancer is an emerging area in drug discovery. Cancer is one of the fatal causes of death globally both in the developed and developing countries.¹ Cancer is the second leading cause of death and the number of new cases is increasing mainly due to environmental prospects.² The World Health Organization

(WHO) stated approximately seven million people deceased from cancer per year and it is estimated that 12 million deaths could occur due to cancer in 2030.³ Nature has been an essential source of new drug candidates since ancient times and a large number of the anticancer agents approved are either natural product resources or their analogues.⁴ Therefore, natural products represent an inspiring resource of drug leads for the designing of new pharmaceuticals with the improved medicinal profile. Hence, there is an urgent need to search for some newer, safer and more effective anticancer agents to improve both life expectancy and quality of a patient's life.

The development of environmentally benign and sustainable methodologies for high value-added chemicals from renewable natural resources has received great interest in various fields of synthetic organic community in both academic and industrial

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research due to recent economic and public health burden views.^{5,6} Among various renewable resources, cashew nut shell liquid (CNSL), low-cost agricultural byproduct and waste in cashew processing industry, is one of the most commonly used renewable resources.⁶ The Food and Agricultural Organization reported that large amounts of cashew nut shell liquid (CNSL), over 0.3 million tons per year, are produced in Asia, Africa, and South America, especially in Brazil, which is a waste and non-edible natural resource.⁷ However, disposal of these industrial feedstock waste to the soil or landfill causes huge environmental problems. To reach these global targets, development of low-cost impact technologies for the utilization of CNSL has become increasingly urgent.

Generally, CNSL contains phenolic derivatives with meta-substituted saturated/unsaturated aliphatic long chain. The major components in CNSL are anacardic acid, cardanol, cardol and 2-methylcardol (Figure 1), in which their composition depends on the basis of the mode of extraction.⁸ CNSL liquid is classified into two types: natural CNSL and technical CNSL. Natural CNSL is obtained by solvent extraction method or cold method, which contains mainly anacardic acid (60-65%), cardanol (10%), cardol (15-20%) and smaller amounts of 2-methyl cardol.⁹ However, technical CNSL obtained through roasting or hot oil process contains cardanol as the main component generated via decarboxylation of anacardic acid. The proportion of CNSL components is cardanol (60-65%), cardol (15-20%) and a smaller amount of methyl cardol (Figure 1).¹⁰

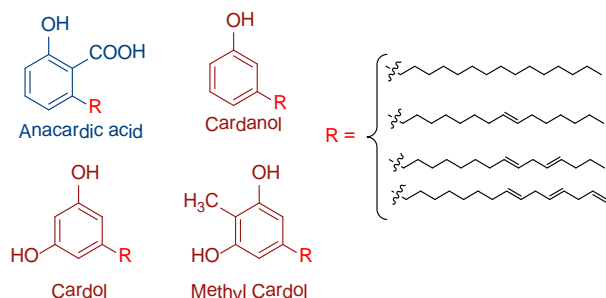


Figure 1. Components of CNSL.

CNSL based components have wide applications in industry as raw material for the fine chemical process, epoxyresins, pesticides, varnishes, adhesives, bio-based polymers, lamination, automobile and as an additive in the surface coating as well as key precursors in pharmaceutical applications.¹¹ CNSL constituents cardanol and anacardic acid derivatives play a diverse role in the field of medicinal chemistry due to its various biological activities, such as antitumor,^{12a} insecticidal,^{12b} enzyme inhibitor,^{12c} antioxidant,^{12d} antibacterial,^{12e} antifungal,^{12f} and larvicide activity.^{12g} The chemical components of CNSL have free phenolic OH groups and double bonds on the side chain attached to a phenolic ring susceptible to chemical modification.¹³ The literature describes many selective chemical modifications on CNSL components, including alkylation, acylation, allylation, hydrogenation, nitration, halogenation, epoxidation, amination, and cyclocarbonylation.¹⁴ The mixture

plays a major role in organic synthesis and it is being used as a building block for the academic and industrial research, and it replaces phenol in many reactions with better results due to its low-cost availability, biodegradability, and renewable natural material.¹⁵ Therefore, based on this versatile chemical structure, much attention has been drawn towards CNSL.

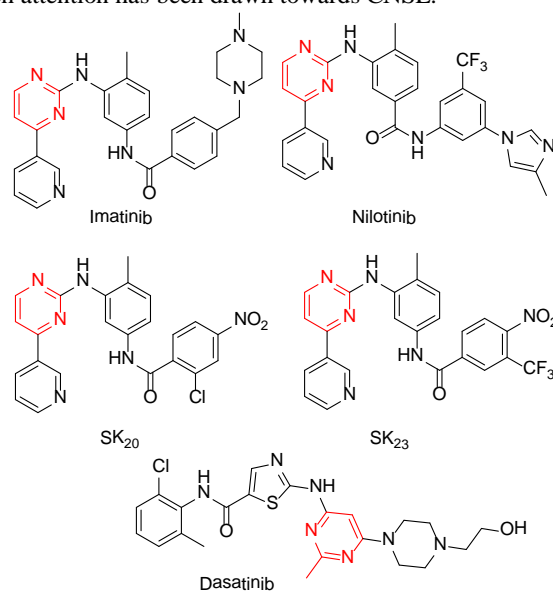


Figure 2. Representative biologically active pyrimidine derivatives.

On the other hand, the structural diversity and biological importance of nitrogen containing heterocycles, including pyrimidine, piperidine, triazole, peptide, and tetrazole have made them attractive targets for synthesis over the years because they are found in many natural products with biological importance and they have been intensively used as versatile scaffolds for drug discovery.¹⁶ Amongst the numerous scaffolds, pyrimidine and their derivatives have received much attention in various fields of synthetic organic chemistry that can be renewed into useful molecules.¹⁷ The pyrimidine scaffold is constituent of a large number of medicines (Figure 2) and can display a broad spectrum of interesting pharmacological and physiological activities, including antiviral,^{18a} antibacterial,^{18b} antifungal,^{18c} anti-filarial,^{18d} antioxidant,^{18e} anthelmintic,^{18f} anticancer,^{18g} anti-HIV,^{18h} antimalarial,¹⁸ⁱ and antitubercular.^{18j} activities. For example Imatinib is a pyrimidine scaffold known as a tyrosine kinase inhibitor and is licensed for the treatment of patients with chronic myeloid leukemia and gastrointestinal stromal tumors.¹⁹ Although many synthetic methods have been developed for the preparation of pyrimidine scaffolds, the development of novel synthetic methods is in great progress.

Morita-Baylis-Hillman reaction is a powerful C-C bond forming method for generating highly functionalized molecules in an atom-economical manner from an activated alkene and a carbonyl compound under the influence of suitable catalyst such as tertiary amine or phosphine.²⁰ Baylis-Hillman adducts holding versatile functionalities i.e. alkene, hydroxyl, and an electron withdrawing functional group in close proximity are valuable structural motifs for diverse organic reactions and transformations.²⁰ These adducts have also been used as

valuable synthons for the architecture of representative natural products, unnatural products, useful heterocycles and bioactive molecules.²¹ Recently, various promising functionalized molecular frameworks derived from Baylis-Hillman reaction and their acetates displayed good biological profile.²²

To the best of our knowledge, no report has yet explored the Baylis-Hillman reaction of cardanol and anacardic acid. On this basis, we found valuable to develop new chemical entities for new drug design utilizing cashew nut shell liquid (CNSL). In continuation of our ongoing efforts endowed with the discovery of potential chemotherapeutic agents,²³ we investigated facile route for the synthesis of novel Baylis-Hillman adducts and their acetates from aldehydes derived from inexpensive cardanol and anacardic acid. Further, based on the concept of molecular hybridization and taken into account the Imatinib molecular framework in the treatment of cancer, we report herein the synthesis of novel pyrimidine probes by using Baylis-Hillman acetates derived from cardanol and anacardic acid.

RESULTS AND DISCUSSION

Synthesis of Baylis-Hillman adducts and acetates from CNSL

Initially, the required cardanol aldehyde (**2**) was synthesized from cardanol ene-mixture (**1**), which was isolated from commercially available cashew nut shell liquid (CNSL) by vacuum distillation according to the reported method.²⁴ Next, cardanol ene-mixture (**1**) was subjected to ozone treatment in the presence of ethyl acetate at $-78\text{ }^{\circ}\text{C}$ for 3 h, followed by reduced with Zn in acetic acid furnished cardanol aldehyde (**2**, C7-CHO) in 78% yield.²⁵ Cardanol aldehyde (**2**) was alkylated using alkyl halides such as methyl iodide, propyl bromide, butyl bromide, pentyl bromide and hexyl bromide in presence of K_2CO_3 in acetone to provide the corresponding alkylated cardanol aldehydes (**3a-3e**) in 83-90% yield (Scheme 1). Anacardic acid ene-mixture (**6**) was obtained from cashew nut by solvent extraction according to the reported procedure.²⁶ After that, the methylated anacardic acid aldehyde (**8**, C7-CHO) was achieved by the same method used to furnish cardanol aldehyde (**2**) as outlined in Scheme 2.²⁵

Initial investigations led to the selection of cardanol aldehyde (**2**) and acrylonitrile (**4a**) as model substrates for optimization of reaction conditions and the results are outlined in Table 1. To select a favorable reaction conditions, we initiated our study by testing the applicability of the catalyst (DABCO, 10 mol%) with a variety of solvents at room temperature. From this screening, we obtained an excellent yield of BH adduct (**5a**) with the reaction performed without any additional solvent at room temperature after 12 h. Therefore, an initial reaction was executed using THF:H₂O (1:1) with acrylonitrile and DABCO (10 mol%) as a catalyst and, the desired product could be obtained in 80% yield at room temperature after 12 h (entry 1). When the reaction was performed using water as a co-solvent with dioxane (1:1) the desired product could be obtained in 90% yield (entry 2). No product was detected when the reaction was run in dioxane, THF, CH_2Cl_2 , CHCl_3 , CH_3CN , MeOH, and EtOH. Only the starting material was recovered from the

reaction after 24 h. Under similar conditions, the reaction was tested with high boiling solvents such as H₂O, DMSO, and DMF to give good yields (70-85%). Interestingly, the complete starting material conversion was observed and BH adduct yield could be improved up to 90% when the reaction was performed without any additional solvent at room temperature, which was chosen as standard reaction condition (entry 13). It should be noted that without a catalyst, no reaction proceeded at all (entry 14). Subsequently, after this optimization, several alkyl substituted cardanol aldehydes (**3a-3e**) and methyl anacardic acid aldehyde (**8**) could be produced into the corresponding BH adducts (**5b-5f** and **9a**) in good yields (Scheme 1 & 2, Table 2).

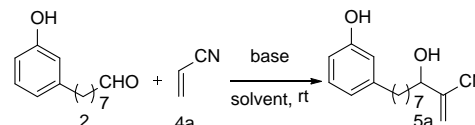


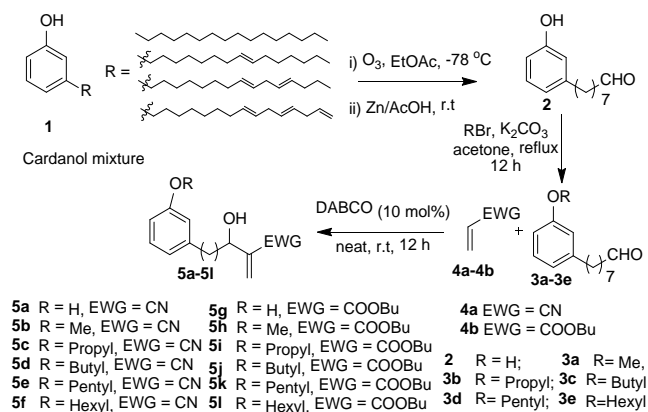
Table 1. Optimization of the reaction conditions.^a

Entry	Solvent	Base	Time (h)	Yield (%) ^b
1	THF:H ₂ O(1:1)	DABCO	24	80
2	Dioxane:H ₂ O(1:1)	DABCO	24	90
3	THF	DABCO	24	- ^c
4	Dioxane	DABCO	24	-
5	MeOH	DABCO	24	-
6	CH_2Cl_2	DABCO	24	-
7	CH_3CN	DABCO	24	-
8	EtOH	DABCO	24	-
9	CHCl_3	DABCO	24	-
10	H ₂ O	DABCO	24	70
11	DMSO	DABCO	24	85
12	DMF	DABCO	24	80
13	Neat	DABCO	12	90
14	Neat	-	12	-

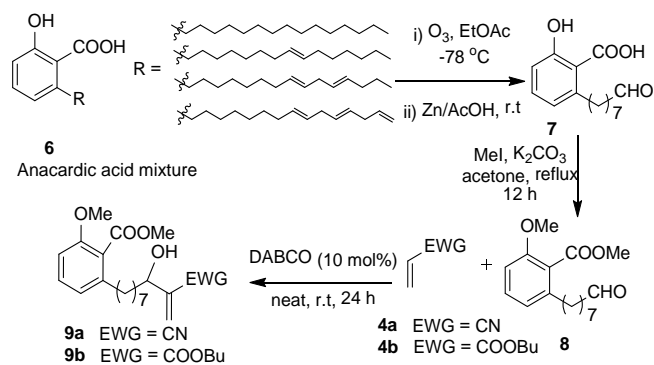
^aReaction conditions: aldehyde **2** (1.0 mmol), acrylonitrile **4a**(1.5 mmol) and DABCO(10 mol%), ^bisolated yield, ^cno reaction.

Further, several cardanol aldehydes (**2**, **3a-3e**) and methyl anacardic acid aldehyde (**8**) were explored with butyl acrylate (**4b**) and DABCO as a catalyst under solvent-free conditions. The cardanol aldehyde (**2**) furnished BH adduct (**5g**) in good yield. In contrast, alkylated aldehydes (**3a-3e**) showed low reactivity towards butyl acrylate (less than 50% conversion). A probable reason could be due to the electronic effect of the alkyl chain on the phenyl ring. To overcome this problem, several modifications were attempted, including prolonged reaction time up to 24 h, the amount of butyl acrylate increased 1.5 to 3 equivalents with respect to aldehyde and PEG(1 mL) as an additive. These modifications made possible to obtain the corresponding BH adducts (**5h-5l**, **9b**) in good yields (Scheme 1 & 2, Table 2).

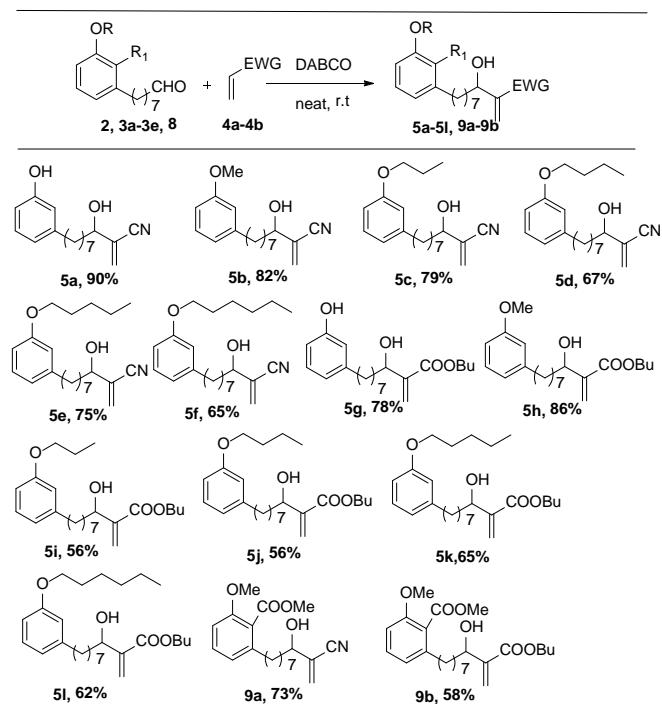
To investigate the scalability of the method, 1.0 g of methyl cardanol aldehyde (**3a**) was used to react with acrylonitrile (**4a**) under standard reaction conditions that provided the BH adduct (**5b**) in good yield (85%).



Scheme 1. Synthesis of Baylis-Hillman adducts from cardanol.

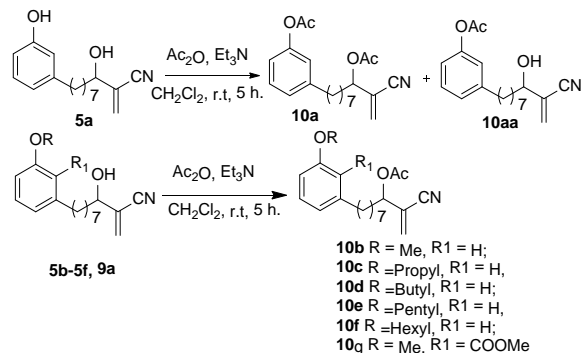


Scheme 2. Synthesis of Baylis-Hillman adducts from anacardic acid.

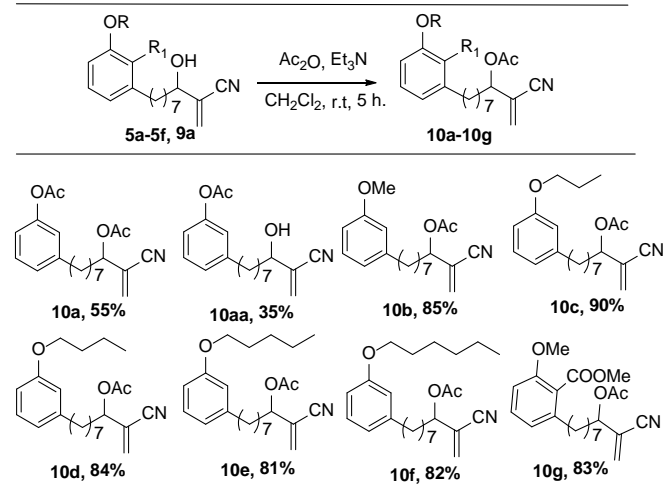
Table 2. Scope and limitations for various Baylis-Hillman adducts.^a

^aReaction conditions: aldehyde (2.0 mmol), alkene (3.0 mmol) and DABCO (10 mol%).

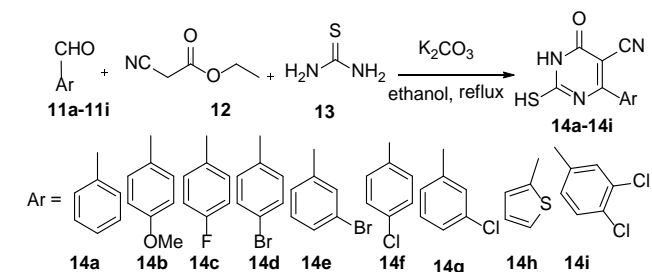
The obtained nitrile BH adducts derived from cardanol and anacardic acid derivatives were easily converted into acetates as shown in Scheme 3 and Table 3. All nitrile BH adducts (**5b-5f**, **9a**) except **5a** were acetylated with acetic anhydride in the presence of triethylamine in CH_2Cl_2 to give the corresponding acetylated compounds (**10b-10g**) as the sole product in excellent yields, whereas in the case of compound **5a** both phenolic and allylic hydroxyl groups could be acetylated. All the synthesized BH adducts and their acetates were well characterized by spectroscopic analysis and further screened for their *in vitro* antitumor activity.



Scheme 3. Synthesis of Baylis-Hillman acetates.

Table 3. Scope and limitations for various Baylis-Hillman acetates.^a

^aReaction conditions: BH adduct (1.0 mmol), Et_3N (2.0 mmol), Ac_2O (2.0 mmol) and CH_2Cl_2 (10 mL).

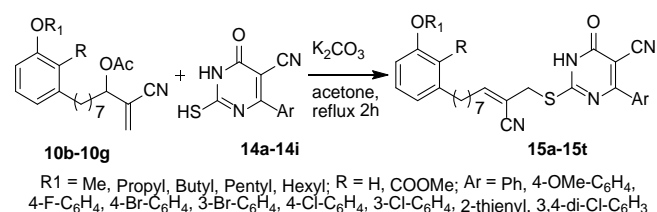


Scheme 4. Synthesis of thiopyrimidine derivatives.

Synthesis of phenolic based pyrimidine derivatives from Baylis-Hillman acetates

Having succeeded in synthesizing Baylis-Hillman alcohols and acetates from cardanol and anacardic acid, we further synthesized phenolic based pyrimidine derivatives. The synthetic strategy for the proposed target phenolic based pyrimidine hybrid scaffolds (**15a-15t**) is outlined in Scheme 5.

At this stage, the aryl-2-sulfanyl-1,6-dihydropyrimidine-5-carbonitriles (**14a-14i**) could be prepared according to the reported procedure as depicted in Scheme 4.²⁷ Cyclisation of aldehydes (**11a-11i**), ethyl cyanoacetate (**12**), and thiourea (**13**) in ethanol as solvent, in the presence of potassium carbonate at reflux temperature for 5 h, furnished the corresponding aryl-2-sulfanyl-1,6-dihydropyrimidine-5-carbonitriles (**14a-14i**) in good yields (Scheme 4). The obtained Baylis-Hillman acetates (**10b-10g**) were treated with various aryl-2-sulfanyl-1,6-dihydropyrimidine-5-carbonitriles (**14a-14i**) using K_2CO_3 in acetone at 40 °C for 2 h, to provide (E)-2-((2-cyano-10-(3-methoxyphenyl)dec-2-en-1-yl)thio)-6-oxo-4-phenyl-1,6-dihydropyrimidine-5-carbonitriles (**15a-15t**) in good yields (Scheme 5 and Table 4).

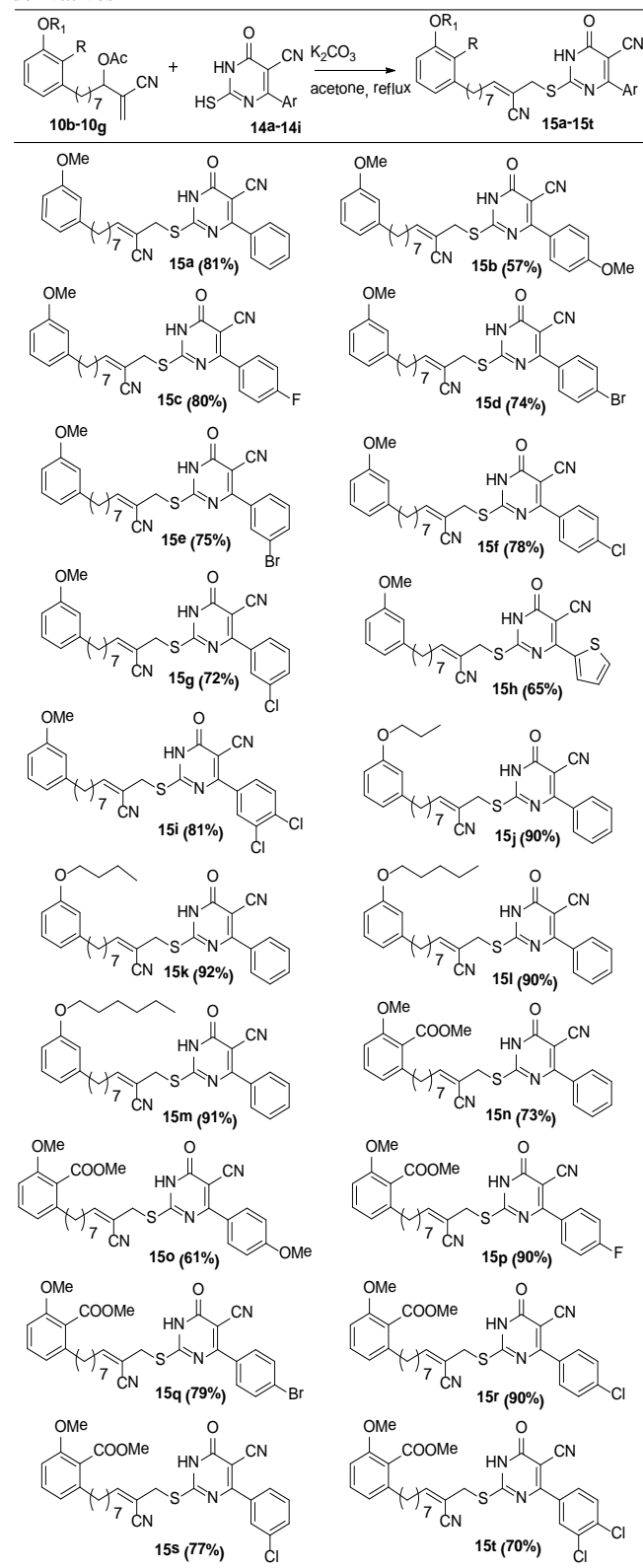


Scheme 5. Synthesis of phenolic based pyrimidine derivatives.

We optimized the reaction conditions with Baylis-Hillman acetate (**10b**) and aryl-2-sulfanyl-1,6-dihydropyrimidine-5-carbonitrile (**14a**) as a model substrate for this conversion using various bases such as piperidine, pyridine, trimethylamine, K_2CO_3 . The findings suggested that K_2CO_3 was one of the best choice for this conversion. This reaction did not proceed in the absence of a base. Different solvent systems such as DMF, THF, CH_2Cl_2 , CH_3CN , acetone, ethanol and toluene could be examined to facilitate this reaction, but only acetone provided excellent results, emerging as one of the best solvents for this conversion. The synthesis of target compounds in excellent yields (81%) was standardized in presence of K_2CO_3 as base and acetone as a solvent at 40 °C for 1 h. Other Baylis-Hillman acetates (**10b-10g**) were reacted with different aryl-2-sulfanyl-1,6-dihydropyrimidine-5-carbonitriles (**14a-14i**) to obtain the desired products in good to excellent yields (Table 4). Both electron-donating (OMe) and electron-withdrawing (F, Cl, Br) groups in phenyl ring of pyrimidine scaffolds could be executed under the standard conditions, leading to corresponding products. This revealed that pyrimidine substrates bearing electron donating groups provided milder yields (**15b** and **15o**) and pyrimidine substrates bearing a halogen atom (F, Cl, or Br) on phenyl ring gave the desired products in good yields (**15c-15g**, **15i** and **15p-15t**). The effect of various alkoxy groups (methyl, propyl, butyl, and hexyl) of the benzene ring in the BH acetate (**15j-15m**) suggested that diverse alkoxy groups could

deliver the desired products, with superior yields for those with the longest length of alkoxy chain (methyl to Hexyl).

Table 4. Scope and limitations for phenolic based pyrimidine derivatives^a



^aReaction conditions: BH acetate (1.0 mmol), thiopyrimidine (1.0 mmol), K_2CO_3 (1.0 mmol) and acetone (15 mL).

Biological activity

The anticancer activities of the above newly synthesized Baylis-Hillman derivatives and phenolic-based pyrimidine hybrid compounds were screened *in vitro* for their cytotoxic effects against four human cancer cell lines MCF-7 (breast cancer), PC-03 (prostate cancer), HEP G2 (liver cancer) and K562 (leukaemia cancer) by using the SRB and MTT assay methods.²⁸ GI₅₀ is the concentration of drug to cause a 50% reduction in the proliferation of cancer cells. Table 5 and 6 shows the values of growth inhibition of 50% (GI₅₀) of all evaluated cells for synthesized hybrid molecules at 48 hours.

Antitumor activity of Baylis-Hillman adducts and acetates

The results in Table 5 revealed that target Baylis-Hillman compounds **5b** and **5h** showed promising activity, whereas remaining compounds showed moderate to low activity against tested cell lines. Compounds **5a-5f**, **9a**, **10aa**, all the BH adducts bear nitrile functionality at the double bond (Table 2). Compound **5b** having methoxy functionality on the phenyl ring showed promising selective anticancer activity against MCF-7 with the GI₅₀ value 19.86 μM and no activity against PC-03, HEP-G2 and K562 (Table 4). Compounds **5g-5l** and **9b**, all the BH adducts possess ester functionality at the double bond (Table 2). Compound **5h**, having methoxy functionality in the phenyl ring, showed promising activity against MCF-7 (with GI₅₀ of 19.72 μM) and moderate activity against HEP G2 (with GI₅₀ of 30.34 μM), while weak activity against PC-03 (97.93 μM) and K562 (69.87 μM) (Table 5). When Baylis-Hillman adducts were converted into corresponding acetates, their activity decreased. Baylis-Hillman acetates (**10a-10g**) possess nitrile functionality at the double bond (Table 3). The results indicated that presence of alkyl groups on the phenyl ring cancel the activity. Compounds **10a**, **10b**, and **10g** displayed moderate activity, while the remaining compounds **10c-10f** showed no activity against the tested cells.

Previous studies also showed that anacardic acid obtained from cashew nuts have antiproliferative activity against prostate cancer cells (TAN) and isobenzofurans designed from anacardic acids show activity against HL-60 cells leukemia, SF295 glioblastoma and, MDA-MB435 melanoma,²⁹ demonstrating the promising anticancer activity of these compounds.

Antitumor activity of phenolic based pyrimidine derivatives

Our results (Table 6) showed that target phenolic based hybrid compounds were active against all tested cell lines. Among them, compounds **15i**, **15j**, and **15l** showed promising activity against HEP-G2 with the GI₅₀ value of 6.17, 2.87 and 4.56 μM respectively, whereas compounds **15j**, **15k**, **15l**, and **15m** showed promising activity against MCF-7 with the GI₅₀ value of 8.51, 8.64, 8.66 and 5.02 μM respectively. Moreover, compound **15m** showed equipotent activity with the GI₅₀ value of 0.25 μM and compound **15j** showed good activity with the GI₅₀ value of 12.86 μM against K562, as compared to the standard doxorubicin (0.04 to 0.25 μM).

The structure-activity relationship (SAR) analysis indicates that the presence of the methoxy group at the phenyl ring of BH

acetate moiety compounds **15a-15i** showed moderate activity (Table 4 & 6). Subsequently, we investigated the effects of methoxy and halogen atoms on the phenyl ring of pyrimidine scaffold, in order to identify some more potent inhibitors.

Table 5. In vitro anticancer activity of Baylis-Hillman derivatives.

Com. no	MCF-7 GI ₅₀ (μM)	PC-03 GI ₅₀ (μM)	HEP G2 GI ₅₀ (μM)	K562 GI ₅₀ (μM)
5a	>100	>100	98.76±0.76	>100
5b	19.86±0.11	>100	>100	>100
5c	91.42±0.69	>100	>100	>100
5d	84.98±0.63	91.05±0.39	>100	97.45±0.19
5e	76.56±0.13	90.24±0.12	>100	>100
5f	78.31±0.56	65.17±0.11	>100	80.66±0.11
5g	73.17±0.90	>100	>100	82.76±0.06
5h	19.72±0.56	97.93±0.72	30.34±0.14	69.87±0.40
5i	71.69±0.64	>100	69.14±0.33	62.60±0.66
5j	>100	>100	>100	>100
5k	>100	>100	>100	>100
5l	>100	>100	>100	>100
9a	75.26±0.71	63.68±0.50	>100	91.30±0.05
9b	59.44±0.78	62.54±0.25	61.82±0.46	99.73±0.12
10a	67.14 ±0.11	>100	81.13±0.52	72.04±0.13
10aa	85.6±0.152	>100	>100	>100
10b	63.74±0.12	78.92±0.21	97.13±0.68	72.18±0.11
10c	>100	>100	>100	>100
10d	>100	>100	>100	>100
10e	>100	>100	>100	>100
10f	>100	>100	>100	>100
10g	>100	>100	64.52±0.41	62.53±0.08
^a	0.07±0.02	0.6±0.02	0.47±0.03	0.25±0.08

^aDoxorubicin is the positive control

Keeping the methoxy group (**15a-15i**) at the phenyl ring of BH acetate moiety but with the presence of methoxyl, halogen groups (F, Cl, and Br) and 2-thienyl on the phenyl ring of pyrimidine scaffold (**15a-15i**) had no significant influence on the activity. Compounds **15a-15h** showed good to moderate activity against all tested cells with GI₅₀ values in the range of 15.95-249.21 μM. In contrast, compound **15i** bearing two chlorine atoms at 3,4 position of the phenyl ring of pyrimidine moiety, showed promising activity against HEP G2 (6.17 μM), and moderate activity against MCF-7 (25.41 μM) and PC-03 (24.98 μM), while less activity against K562 (170.11 μM).

Interestingly, the methyl group on the phenyl ring of BH acetate when replaced by propyl, butyl, pentyl and hexyl groups (**15j-15m**) in absence of any substitution on the phenyl ring of pyrimidine scaffold confer an increased activity. Compound **15j** bearing propyl group on phenyl ring showed promising activity against MCF-7 (8.51 μM) and HEP-G2 (2.87 μM), while moderate activity against PC-03 (23.71 μM) and good activity against K562 (12.86 μM). Compound **15k** having butyl group on phenyl ring showed promising activity against MCF-7 (8.64 μM), and moderate activity against for PC-03 (36.17 μM), HEP-G2 (15.24 μM) and K562 (24.45 μM). Compound **15l** having pentyl group on phenyl ring showed promising activity against MCF-7 (8.66 μM) and HEP-G2 (4.56 μM), whereas it showed

moderate activity against PC-03 (22.62 μM) and K562 (24.62 μM). It can be verified that compound **15m** having hexyl group on phenyl ring showed substantial activity for K562 (0.25 μM) and promising activity against MCF-7 (5.02 μM), however, showed moderate activity against PC-03 (18.36 μM) and HEP-G2 (22.42 μM). These findings clearly indicate that the compound **15m** was the most potent compound. Thus, the length of the carbon chain of phenyl ring played an important role for the activity (Table 6).

Table 6. *In vitro* anticancer activity data of compounds **15a-15t**

Comp. no	MCF-7 GI ₅₀ (μM)	PC-03 GI ₅₀ (μM)	HEP G2 GI ₅₀ (μM)	K562 GI ₅₀ (μM)
15a	26.64±0.33	71.19±0.29	28.15±0.20	>100
15b	30.41±0.40	25.67±1.32	>100	>100
15c	27.13±0.45	25.40±0.39	23.73±0.88	>100
15d	58.22±0.31	25.70±0.43	78.81±0.96	24.49±0.06
15e	16.12±0.41	26.14±0.89	35.35±3.14	24.28±0.55
15f	31.28±0.21	25.95±0.46	19.01±0.32	>100
15g	27.34±0.45	26.23±0.67	58.60±0.52	23.99±0.12
15h	24.24±0.30	24.00±0.27	15.95±0.16	24.59±0.22
15i	25.41±0.90	24.98±0.51	6.17±0.31	>100
15j	8.51±0.45	23.71±0.65	2.87±0.47	12.86±0.42
15k	8.64±0.33	36.17±0.82	15.24±0.62	24.45±0.18
15l	8.66±0.44	22.62±0.42	4.56±0.68	24.62±0.10
15m	5.02±0.23	18.36±0.56	22.42±1.13	0.25±0.06
15n	26.25±0.51	>100	30.72±2.21	>100
15o	22.21±0.37	24.34±0.43	24.79±0.39	>100
15p	27.02±0.32	29.58±0.37	25.06±0.94	35.44±0.09
15q	22.28±0.43	25.17±2.10	14.04±0.27	25.95±0.61
15r	28.58±0.51	25.40±0.72	45.10±0.44	>100
15s	>100	>100	>100	>100
15t	27.02±0.42	51.03±2.3	28.21±0.74	35.44±0.13
^a	0.07±0.02	0.6±0.02	0.47±0.03	0.25±0.08

^aDoxorubicin is the positive control.

The **Figure 3** summarizes the *in vitro* antitumor activity of main compounds found in this study.

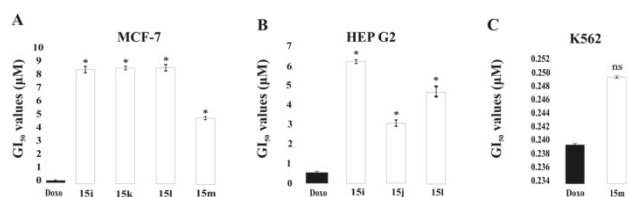


Figure 3: *In vitro* anticancer activity of compounds **15i-15m**. The bar graphs show the concentration in μM of each compound required for 50% inhibition (GI₅₀) of MCF-7, HEP G2 and K562 cell lines compared to Doxorubicin (positive control). **Note:** * $p < 0.05$ was considered as significant and ns (no significant).

Further modifications including compounds bearing methoxy group and methyl ester (COOMe) groups (**15n-15t**) on the phenyl ring of BH acetate and the presence of methoxy group and halogen groups (F, Cl and Br) on the phenyl ring of pyrimidine scaffold had no much influence on the anticancer

activity against all tested cell lines, while compound **15s** having chlorine at meta position of phenyl ring of pyrimidine scaffold led to dramatically decreasing the activity. Compounds **15n-15t** showed moderate to small activities towards all tested cell lines with GI₅₀ values in the range of 14.04->250 μM (Table 6).

EXPERIMENTAL

General remarks

All reagents were analytical grade and used without further purification. Chromatographic purification was performed on silica gel (Merck, 100-200 mesh) and analytical thin layer chromatography (TLC) on silica gel 60-F₂₅₄. ¹H NMR (300 MHz) and ¹³C NMR (75 MHz) spectra were measured with a Bruker Avance DPXT-300 spectrometer with CDCl₃ and DMSO-d₆ as solvents and recorded in ppm relative to internal tetramethylsilane standard (TMS). The ¹H NMR spectra are reported as follows: ppm (multiplicity, coupling constant J/Hz, number of protons). Multiplicity is abbreviated as follows: s (singlet), d (doublet), dd (doublet of doublets), t (triplet), m (multiplet). Coupling constants (*J*) are quoted in Hertz and recorded to the nearest 0.1 Hz. High-resolution mass spectroscopy (HRMS) was performed on a UFLC Shimadzu LC-20AD apparatus, with and IES-Q-QTOF-microTOF III detector (Bruker Daltonics) in chemical ionization positive ion mode (*m/z* 120-1200). The samples were prepared with 0.1 g/mL (methanol/ water 7:3) and injected 1 μL , using elution gradient water (phase A) and acetonitrile (phase B), both with acetic acid 1%, isocratic method 50% and the running time of 3 min. The infrared spectra were recorded on a FT-IR spectrometer and reported as wavenumbers (cm⁻¹).

General experimental procedure for the synthesis of Baylis-Hillman adducts (**5a-5l**, **9a-9b**)

Several substituted aldehydes (**2**, **3a-3e**, **8**) (2 mmol), activated olefin (**4a-4b**) (3 mmol) and DABCO (10 mol% with respect to aldehyde) were mixed well and allowed to stir at room temperature until reaction is completed (reaction monitored by the TLC). (In case of aldehydes **3a-3e**, **8** with butyl acrylate **4b** showed less reactivity. Then reaction time was increased to 12-24 h, butyl acrylate **4b** (6 mmol) and PEG (1 mL) as additive). After completion, the reaction mixture was diluted with water (15 mL) and extracted with ethyl acetate (3x20 mL). The combined organic layers were dried over sodium sulphate, filtered, concentrated under vacuum to give the crude product. The product was purified by silica gel column chromatography using ethyl acetate:hexane as eluent (2:8), providing corresponding BH adducts (**5a-5l**, **9a-9b**) in good yield as colorless liquids.

3-Hydroxy-10-(3-hydroxyphenyl)-2-methylenedecanenitrile

(**5a**): Colorless liquid; ¹H NMR (300 MHz, CDCl₃): δ 7.14-7.07 (m, 1H), 6.73-6.69 (m, 1H), 6.65-6.61 (m, 2H), 5.97-5.96 (m, 1H), 5.96-5.95 (m, 1H), 5.66 (s, 1H), 4.24-4.16 (m, 1H), 2.52 (t, *J* = 7.3 Hz, 2H), 1.94 (s, 1H), 1.74-1.50 (m, 4H), 1.34-1.22 (m, 8H); ¹³C NMR (75 MHz, CDCl₃): δ 165.2, 155.5, 144.6, 130.0, 129.3, 126.7, 120.7, 115.3, 112.5, 72.3, 35.6, 35.5, 30.9, 29.0, 28.9, 28.8, 24.8; IR (neat): 3386, 3039, 2927, 2854, 2229, 1589,

1485, 1458, 1269, 1234, 1157, 948 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{17}\text{H}_{24}\text{NO}_2$: 274.1807 $[\text{M}+\text{H}]^+$; found. 274.1813.

3-Hydroxy-10-(3-methoxyphenyl)-2-methylenedecanenitrile (5b): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.23-7.16 (m, 1H), 6.77 (d, $J = 7.6$ Hz, 1H), 6.74-6.70 (m, 2H), 6.00-5.96 (m, 2H), 4.23 (t, $J = 6.7$ Hz, 1H), 3.79 (s, 3H), 2.57 (t, $J = 7.3$ Hz, 2H), 1.77-1.55 (m, 4H), 1.42-1.28 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 159.4, 144.3, 129.6, 129.0, 126.9, 120.7, 116.9, 114.1, 110.7, 72.2, 55.0, 35.8, 35.5, 31.1, 29.1, 29.0(2), 24.9; IR (neat): 3459, 2997, 2931, 2854, 2225, 1600, 1585, 1488, 1465, 1454, 1434, 1315, 1261, 1180, 1045, 948 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{18}\text{H}_{26}\text{NO}_2$: 288.1964 $[\text{M}+\text{H}]^+$; found. 288.1950.

3-Hydroxy-2-methylene-10-(3-propoxyphenyl)decanenitrile (5c): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.21-7.14 (m, 1H), 6.78-6.69 (m, 3H), 5.99-5.95 (m, 2H), 4.25-4.19 (q, $J = 5.7$ Hz, 1H), 3.91 (t, $J = 6.5$ Hz, 2H), 2.57 (t, $J = 7.4$ Hz, 2H), 2.13 (d, $J = 5.1$ Hz, 1H), 1.86-1.56 (m, 6H), 1.43-1.28 (m, 8H), 1.03 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 159.0, 144.3, 129.6, 129.0, 126.9, 120.6, 116.9, 114.7, 111.3, 72.3, 69.3, 35.9, 35.6, 31.2, 29.2, 29.0, 24.9, 22.5, 10.5; IR (neat): 3471, 3031, 2958, 2931, 2854, 2225, 1600, 1581, 1485, 1450, 1392, 1261, 1157, 1049, 948 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{20}\text{H}_{30}\text{NO}_2$: 316.2277 $[\text{M}+\text{H}]^+$; found. 316.2272.

10-(3-Butoxyphenyl)-3-hydroxy-2-methylenedecanenitrile (5d): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.22-7.14 (m, 1H), 6.78-6.69 (m, 3H), 6.00-5.95 (m, 2H), 4.26-4.17 (m, 1H), 3.95 (t, $J = 6.4$ Hz, 2H), 2.57 (t, $J = 7.4$ Hz, 2H), 2.30-2.20 (m, 1H), 1.82-1.69 (m, 4H), 1.66-1.57 (m, 2H), 1.55-1.44 (m, 2H), 1.40-1.27 (m, 8H), 0.98 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 159.0, 144.3, 129.7, 129.0, 126.9, 120.6, 117.0, 114.7, 111.3, 72.2, 67.4, 35.8, 35.5, 31.3, 31.2, 29.2, 29.0, 24.9, 19.2, 13.8; IR (neat): 3463, 3031, 2931, 2854, 2225, 1600, 1581, 1488, 1450, 1392, 1311, 1257, 1157, 1068, 1029, 945 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{21}\text{H}_{32}\text{NO}_2$: 330.2433 $[\text{M}+\text{H}]^+$; found. 330.2429.

3-Hydroxy-2-methylene-10-(3-pentyloxyphenyl)decanenitrile (5e): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.21-7.14 (m, 1H), 6.77-6.69 (m, 3H), 5.99-5.95 (m, 2H), 4.26-4.18 (q, $J = 5.2$ Hz, 1H), 3.94 (t, $J = 6.5$ Hz, 2H), 2.56 (t, $J = 7.4$ Hz, 2H), 2.10 (d, $J = 5.2$ Hz, 1H), 1.83-1.55 (m, 6H), 1.49-1.26 (m, 12H), 0.93 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 159.0, 144.3, 129.6, 129.0, 126.9, 120.6, 116.9, 114.7, 111.3, 72.3, 67.8, 35.9, 35.6, 31.2, 29.2, 29.0, 28.9, 28.1, 24.9, 22.4, 13.9; IR (neat): 3456, 3031, 2931, 2854, 2225, 1600, 1581, 1488, 1465, 1450, 1392, 1284, 1261, 1157, 1076, 1053, 1033, 945 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{22}\text{H}_{34}\text{NO}_2$: 344.2590 $[\text{M}+\text{H}]^+$; found. 344.2590.

10-(3-(Hexyloxy)phenyl)-3-hydroxy-2-methylenedecanenitrile (5f): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.21-7.13 (m, 1H), 6.77-6.66 (m, 3H), 5.99-5.95 (m, 2H), 4.25-4.17 (m, 1H), 3.94 (t, $J = 6.5$ Hz, 2H), 2.56 (t, $J = 7.6$ Hz, 2H), 2.17-2.11 (m, 1H), 1.83-1.54 (m, 6H), 1.50-1.22 (m, 14H), 0.91 (t, $J = 6.7$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 159.1, 144.3, 129.7, 129.1, 126.9, 120.6, 117.0, 114.8, 111.4, 72.3, 67.8, 35.9, 35.6, 31.6, 31.2, 29.3, 29.1, 25.7,

25.0, 22.6, 14.0; IR (neat): 3459, 3031, 2927, 2854, 2225, 1600, 1581, 1488, 1465, 1450, 1392, 1311, 1261, 1157, 1045, 948 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{23}\text{H}_{36}\text{NO}_2$: 358.2746 $[\text{M}+\text{H}]^+$; found. 358.2733.

Butyl 3-hydroxy-10-(3-hydroxyphenyl)-2-methylenedecanoate (5g): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.16-7.09 (m, 1H), 6.73 (d, $J = 7.3$ Hz, 1H), 6.67-6.62 (m, 2H), 6.21 (s, 1H), 5.77 (s, 1H), 5.22 (brs, 1H), 4.43-4.34 (m, 1H), 4.18 (t, $J = 6.5$ Hz, 2H), 2.67 (brs, 1H), 2.54 (t, $J = 7.3$ Hz, 2H), 1.72-1.53 (m, 6H), 1.48-1.22 (m, 10H), 0.95 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 166.8, 155.7, 144.6, 142.4, 129.2, 124.8, 120.5, 115.3, 112.5, 71.8, 64.8, 36.1, 35.6, 31.0, 30.5, 28.9, 25.6, 19.1, 13.6; IR (neat): 3390, 2927, 2854, 1697, 1623, 1589, 1485, 1458, 1272, 1157, 1095, 1064, 952 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{21}\text{H}_{32}\text{O}_4\text{Na}$: 371.2198 $[\text{M}+\text{Na}]^+$; found. 371.2186.

Butyl 3-hydroxy-10-(3-methoxyphenyl)-2-methylenedecanoate (5h): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.22-7.15 (m, 1H), 6.79-6.69 (m, 3H), 6.20 (s, 1H), 5.76 (s, 1H), 4.40-4.33 (q, $J = 6.5$ Hz, 1H), 4.18 (t, $J = 6.5$ Hz, 2H), 3.79 (s, 3H), 2.60-2.53 (m, 3H), 1.72-1.55 (m, 8H), 1.48-1.26 (m, 8H), 0.95 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 166.6, 159.5, 144.5, 142.7, 129.1, 124.5, 120.8, 114.1, 110.7, 71.8, 64.6, 55.0, 36.2, 35.9, 31.3, 30.5, 29.4, 29.3, 29.2, 25.8, 13.6; IR (neat): 3444, 2927, 2854, 1712, 1600, 1585, 1488, 1458, 1434, 1261, 1153, 1099, 1045, 952 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{22}\text{H}_{34}\text{O}_4\text{Na}$: 385.2355 $[\text{M}+\text{Na}]^+$; found: 385.2342.

Butyl 3-hydroxy-2-methylene-10-(3-propoxyphenyl)decanoate (5i): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.20-7.13 (m, 1H), 6.77-6.68 (m, 3H), 6.20 (s, 1H), 5.76 (s, 1H), 4.40-4.34 (q, $J = 6.2$ Hz, 1H), 4.18 (t, $J = 6.5$ Hz, 2H), 3.90 (t, $J = 6.5$ Hz, 2H), 2.61 (d, $J = 6.2$ Hz, 1H), 2.56 (t, $J = 7.6$ Hz, 2H), 1.86-1.74 (m, 2H), 1.72-1.55 (m, 6H), 1.48-1.26 (m, 10H), 1.03 (t, $J = 7.4$ Hz, 3H), 0.95 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 166.6, 159.0, 144.3, 142.6, 129.0, 124.5, 120.5, 114.7, 111.3, 71.7, 69.2, 64.6, 36.1, 35.9, 31.2, 30.5, 29.3, 29.2, 25.7, 22.6, 19.1, 13.5, 10.4; IR (neat): 3494, 2958, 2931, 2854, 1712, 1600, 1581, 1450, 1392, 1261, 1157, 1064, 952 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{24}\text{H}_{38}\text{O}_4\text{Na}$: 413.2688 $[\text{M}+\text{Na}]^+$; found: 413.2667.

Butyl 10-(3-butoxyphenyl)-3-hydroxy-2-methylenedecanoate (5j): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.20-7.12 (m, 1H), 6.77-6.67 (m, 3H), 6.21 (s, 1H), 5.76 (s, 1H), 4.41-4.34 (q, $J = 6.7$ Hz, 1H), 4.18 (t, $J = 6.5$ Hz, 2H), 3.94 (t, $J = 6.4$ Hz, 2H), 2.61 (d, $J = 6.7$ Hz, 1H), 2.56 (t, $J = 7.6$ Hz, 2H), 1.81-1.56 (m, 8H), 1.54-1.26 (m, 12H), 1.02-0.91 (m, 6H); ^{13}C NMR (75 MHz, CDCl_3): δ 166.6, 159.1, 144.4, 129.0, 124.5, 120.6, 114.7, 111.3, 71.8, 67.4, 64.6, 36.2, 35.9, 31.3, 31.3, 30.5, 29.34, 29.3, 29.2, 25.8, 19.2, 19.1, 13.8, 13.6; IR (neat): 3467, 3031, 2958, 2858, 1712, 1627, 1600, 1581, 1485, 1458, 1392, 1261, 1157, 1068, 1029, 952 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{25}\text{H}_{41}\text{O}_4$: 405.3005 $[\text{M}+\text{H}]^+$; found: 405.2996.

Butyl 3-hydroxy-2-methylene-10-(3-pentyloxyphenyl)decanoate (5k): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.21-7.13 (m, 1H), 6.80-6.68 (m, 3H),

6.21 (s, 1H), 5.76 (s, 1H), 4.43-4.34 (m, 1H), 4.18 (t, $J = 6.5$ Hz, 2H), 3.93 (t, $J = 6.5$ Hz, 2H), 2.62 (d, $J = 6.7$ Hz, 1H), 2.56 (t, $J = 7.3$ Hz, 2H), 1.84-1.56 (m, 8H), 1.50-1.28 (m, 14H), 1.03-0.90 (m, 6H); ^{13}C NMR (75 MHz, CDCl_3): δ 166.6, 159.1, 144.4, 142.7, 129.0, 124.5, 120.6, 114.7, 111.3, 71.8, 67.7, 64.6, 36.2, 35.9, 31.3, 30.5, 29.4, 29.3, 29.2, 29.0, 28.2, 25.8, 22.4, 19.1, 13.9, 13.6; IR (neat): 3490, 3031, 2954, 2931, 2858, 1712, 1600, 1581, 1488, 1465, 1392, 1261, 1157, 1056, 1033, 952 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{26}\text{H}_{43}\text{O}_4$: 419.3161 $[\text{M}+\text{H}]^+$; found: 419.3164.

Butyl 10-(3-(hexyloxy)phenyl)-3-hydroxy-2-methylenedecanoate (5I): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.20-7.13 (m, 1H), 6.77-6.67 (m, 3H), 6.21 (s, 1H), 5.76 (s, 1H), 4.41-4.34 (q, $J = 6.4$ Hz, 1H), 4.18 (t, $J = 6.5$ Hz, 2H), 3.94 (t, $J = 6.4$ Hz, 2H), 2.63 (d, $J = 6.4$ Hz, 1H), 2.56 (t, $J = 7.7$ Hz, 2H), 1.83-1.54 (m, 8H), 1.51-1.26 (m, 16H), 0.98-0.88 (m, 6H); ^{13}C NMR (75 MHz, CDCl_3): δ 166.6, 159.1, 144.4, 142.7, 129.0, 124.5, 120.6, 114.7, 111.3, 71.8, 67.7, 64.6, 36.2, 35.9, 31.5, 31.3, 30.5, 29.4, 29.3, 29.28, 29.2, 25.8, 25.7, 22.5, 19.1, 13.9, 13.6; IR (neat): 3421, 2954, 2927, 2854, 1712, 1600, 1581, 1488, 1461, 1392, 1261, 1157, 1064, 952 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{27}\text{H}_{45}\text{O}_4$: 433.3318 $[\text{M}+\text{H}]^+$; found: 433.3318.

Methyl 2-(9-cyano-8-hydroxydec-9-enyl)-6-methoxybenzoate (9a): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.26 (t, $J = 8.1$ Hz, 1H), 6.81 (d, $J = 7.6$ Hz, 1H), 6.76 (d, $J = 8.1$ Hz, 1H), 5.98 (s, 1H), 5.96 (s, 1H), 4.22 (t, $J = 6.4$ Hz, 1H), 3.90 (s, 3H), 3.81 (s, 3H), 2.53 (t, $J = 7.7$ Hz, 2H), 1.79-1.50 (m, 4H), 1.45-1.24 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.0, 156.1, 141.1, 130.2, 129.6, 129.5, 126.9, 121.4, 117.0, 108.3, 72.1, 55.8, 52.1, 35.5, 33.3, 30.8, 29.1, 29.0, 28.9, 24.8; IR (neat): 3471, 3000, 2931, 2854, 2225, 1731, 1585, 1469, 1434, 1269, 1188, 1110, 1072, 952 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{20}\text{H}_{27}\text{NO}_4\text{Na}$: 368.1838 $[\text{M}+\text{Na}]^+$; found: 368.1846.

Methyl 2-(9-(butoxycarbonyl)-8-hydroxydec-9-enyl)-6-methoxybenzoate (9b): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.30-7.23 (m, 1H), 6.81 (d, $J = 7.6$ Hz, 1H), 6.75 (d, $J = 8.3$ Hz, 1H), 6.20 (s, 1H), 5.76 (s, 1H), 4.41-4.33 (m, 1H), 4.17 (t, $J = 6.5$ Hz, 2H), 3.89 (s, 3H), 3.81 (s, 3H), 2.60 (d, $J = 7.3$ Hz, 1H), 2.52 (t, $J = 7.7$ Hz, 2H), 1.72-1.52 (m, 6H), 1.47-1.25 (m, 10H), 0.94 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 168.8, 166.6, 156.1, 142.7, 141.2, 130.1, 124.4, 121.42, 121.4, 108.3, 71.6, 64.6, 55.7, 52.0, 36.2, 33.3, 31.0, 30.5, 29.3, 29.23, 29.2, 25.7, 19.1, 13.6; IR (neat): 3513, 2931, 2858, 1731, 1716, 1627, 1585, 1469, 1434, 1396, 1269, 1164, 1110, 1087, 956 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{24}\text{H}_{37}\text{O}_6$: 421.2590 $[\text{M}+\text{H}]^+$; found: 421.2592.

General experimental procedure for the synthesis of Baylis-Hillman acetates (10a-10g)

To a well stirred solution of Baylis-Hillmann adduct (**5a-5f**, **9a**) (1 mmol) in dichloromethane (10 mL) was added Et_3N (2 mmol) at 0°C under N_2 atmosphere, then the reaction mixture was stirred for about 15 min. Acetic anhydride (2 mmol) was added slowly at the same temperature and allowed to stir at room temperature until complete consumption of starting

material as monitored by the TLC. After completion, the reaction mixture was diluted with water (10 ml) and extracted with dichloromethane (2x15 mL), then separated the layers and dried over Na_2SO_4 , filtered and concentrated under reduced. The crude product was submitted to silica gel column chromatography using ethyl acetate:hexane as eluent (1:9), providing the corresponding BH acetates (**10a-10g**) as a colorless liquid. The synthesized BH acetates were characterized by spectroscopic techniques.

3-(8-Acetoxy-9-cyanodec-9-enyl)phenyl acetate (10a): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.24-7.20 (m, 1H), 7.00 (d, $J = 7.8$ Hz, 1H), 6.88-6.83 (m, 2H), 5.99 (s, 1H), 5.93 (s, 1H), 5.21 (t, $J = 7.0$ Hz, 1H), 2.56 (t, $J = 7.4$ Hz, 2H), 2.25 (s, 3H), 2.06 (s, 3H), 1.80-1.66 (m, 2H), 1.62-1.51 (m, 2H), 1.31-1.21 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.8, 169.5, 150.5, 144.4, 132.5, 129.0, 125.8, 122.7, 121.3, 118.6, 116.1, 73.1, 35.5, 32.7, 30.9, 29.0, 28.9, 28.8, 24.7, 21.0, 20.8; IR (neat): 2931, 2858, 2225, 1766, 1747, 1612, 1585, 1485, 1442, 1369, 1211, 1118, 1018, 952 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{21}\text{H}_{28}\text{NO}_4$: 358.2018 $[\text{M}+\text{H}]^+$; found: 358.2016.

3-(9-Cyano-8-hydroxydec-9-enyl)phenyl acetate (10aa): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.24-7.20 (m, 1H), 7.00 (d, $J = 7.7$ Hz, 1H), 6.87-6.83 (m, 2H), 5.95-5.92 (m, 2H), 4.20-4.15 (m, 1H), 2.56 (t, $J = 7.6$ Hz, 2H), 2.25 (s, 3H), 1.80 (brs, 1H), 1.70-1.51 (m, 4H), 1.34-1.21 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.7, 150.5, 144.4, 129.7, 129.6, 129.0, 126.9, 125.9, 121.3, 118.6, 72.1, 35.5, 30.9, 29.0, 28.9, 28.8, 24.8, 21.1; IR (neat): 3463, 2927, 2854, 2225, 1766, 1739, 1612, 1585, 1485, 1446, 1369, 1211, 1141, 1014, 948 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{19}\text{H}_{26}\text{NO}_3$: 316.1913 $[\text{M}+\text{H}]^+$; found: 316.1912.

2-Cyano-10-(3-methoxyphenyl)dec-1-en-3-yl acetate (10b): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.22-7.16 (m, 1H), 6.79-6.70 (m, 3H), 6.03 (s, 1H), 5.97 (s, 1H), 5.25 (t, $J = 6.8$ Hz, 1H), 3.79 (s, 3H), 2.57 (t, $J = 7.6$ Hz, 2H), 2.10 (s, 3H), 1.84-1.70 (m, 2H), 1.64-1.54 (m, 2H), 1.36-1.26 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.8, 159.5, 144.3, 132.5, 129.1, 122.8, 120.7, 116.1, 114.1, 110.7, 73.2, 55.0, 35.8, 32.7, 31.2, 29.1, 29.0, 28.9, 24.7, 20.9; IR (neat): 2931, 2854, 2225, 1747, 1600, 1585, 1488, 1458, 1434, 1373, 1261, 1230, 1153, 1045, 1029, 956 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{20}\text{H}_{28}\text{NO}_3$: 330.2069 $[\text{M}+\text{H}]^+$; found: 330.2065.

2-Cyano-10-(3-propoxyphenyl)dec-1-en-3-yl acetate (10c): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.21-7.12 (m, 1H), 6.77-6.64 (m, 3H), 6.03 (s, 1H), 5.97 (s, 1H), 5.25 (t, $J = 6.8$ Hz, 1H), 3.90 (t, $J = 6.5$ Hz, 2H), 2.56 (t, $J = 7.6$ Hz, 2H), 2.10 (s, 3H), 1.86-1.70 (m, 4H), 1.63-1.54 (m, 2H), 1.35-1.21 (m, 8H), 1.03 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.8, 159.0, 144.2, 132.5, 129.0, 122.8, 120.5, 116.1, 114.7, 111.3, 73.1, 69.2, 35.8, 32.7, 31.1, 29.1, 29.0, 28.9, 24.7, 22.5, 20.8, 10.4; IR (neat): 3031, 2931, 2854, 2225, 1747, 1600, 1581, 1488, 1450, 1373, 1261, 1230, 1157, 1049, 1026 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{22}\text{H}_{32}\text{NO}_3$: 358.2382 $[\text{M}+\text{H}]^+$; found: 358.2380.

10-(3-Butoxyphenyl)-2-cyanodec-1-en-3-yl acetate (10d): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.20-7.13 (m,

1H), 6.77-6.58 (m, 3H), 6.03 (s, 1H), 5.97 (s, 1H), 5.26 (t, $J = 6.7$ Hz, 1H), 3.95 (t, $J = 6.4$ Hz, 2H), 2.56 (t, $J = 7.3$ Hz, 2H), 2.10 (s, 3H), 1.83-1.71 (m, 4H), 1.65-1.56 (m, 2H), 1.53-1.45 (m, 2H), 1.38-1.26 (m, 8H), 0.97 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.8, 159.1, 144.2, 132.5, 129.0, 122.8, 120.5, 116.1, 114.7, 111.3, 73.2, 67.4, 35.9, 32.7, 31.3, 31.2, 29.1, 29.0, 28.9, 24.7, 20.8, 19.2, 13.8; IR (neat): 3031, 2858, 2225, 1747, 1600, 1581, 1488, 1450, 1373, 1257, 1230, 1157, 1064, 1026, 956 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{23}\text{H}_{34}\text{NO}_3$: 372.2539 $[\text{M}+\text{H}]^+$; found: 372.2538.

2-Cyano-10-(3-(pentyloxy)phenyl)dec-1-en-3-yl acetate (10e): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.21-7.14 (m, 1H), 6.77-6.68 (m, 3H), 6.03 (s, 1H), 5.97 (s, 1H), 5.25 (t, $J = 6.8$ Hz, 1H), 3.94 (t, $J = 6.5$ Hz, 2H), 2.56 (t, $J = 7.4$ Hz, 2H), 2.10 (s, 3H), 1.83-1.72 (m, 4H), 1.65-1.56 (m, 2H), 1.49-1.38 (m, 4H), 1.36-1.28 (m, 8H), 0.93 (t, $J = 7.01$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.8, 159.1, 144.2, 132.5, 129.0, 122.8, 120.5, 116.1, 114.7, 111.3, 73.2, 67.7, 35.9, 32.7, 31.2, 29.1, 29.0, 28.99, 28.9, 28.1, 24.7, 22.4, 20.8, 13.9; IR (neat): 3031, 2931, 2858, 2225, 1747, 1600, 1581, 1488, 1461, 1450, 1373, 1257, 1230, 1157, 1049, 1026, 952 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{24}\text{H}_{36}\text{NO}_3$: 386.2695 $[\text{M}+\text{H}]^+$; found: 386.2694.

2-Cyano-10-(3-(hexyloxy)phenyl)dec-1-en-3-yl acetate (10f): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.13-7.05 (m, 1H), 6.70-6.58 (m, 3H), 5.95 (s, 1H), 5.89 (s, 1H), 5.18 (t, $J = 6.7$ Hz, 1H), 3.86 (t, $J = 6.4$ Hz, 2H), 2.49 (t, $J = 7.6$ Hz, 2H), 2.02 (s, 3H), 1.77-1.62 (m, 4H), 1.58-1.47 (m, 2H), 1.44-1.34 (m, 2H), 1.31-1.19 (m, 12H), 0.83 (t, $J = 6.5$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.8, 159.1, 144.2, 132.5, 129.0, 122.8, 120.5, 116.1, 114.7, 111.3, 73.2, 67.7, 35.9, 32.7, 31.5, 31.2, 29.2, 29.1, 29.0, 28.9, 25.7, 24.7, 22.5, 20.8, 13.9; IR (neat): 2931, 2858, 2225, 1747, 1600, 1581, 1488, 1450, 1373, 1257, 1230, 1157, 1026, 952 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{25}\text{H}_{38}\text{NO}_3$: 400.2852 $[\text{M}+\text{H}]^+$; found: 400.2834.

Methyl 2-(8-acetoxy-9-cyanodec-9-enyl)-6-methoxybenzoate (10g): Colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.26 (t, $J = 8.3$, 7.6 Hz, 1H), 6.81 (d, $J = 7.6$ Hz, 1H), 6.76 (d, $J = 8.3$ Hz, 1H), 6.03 (s, 1H), 5.97 (s, 1H), 5.25 (t, $J = 7.1$ Hz, 1H), 3.90 (s, 3H), 3.81 (s, 3H), 2.53 (t, $J = 7.6$ Hz, 2H), 2.10 (s, 3H), 1.84-1.70 (m, 2H), 1.62-1.51 (m, 2H), 1.36-1.22 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.8, 168.9, 156.2, 141.1, 132.5, 130.2, 130.1, 122.8, 121.4, 116.1, 108.4, 73.2, 55.8, 52.1, 33.3, 32.8, 30.9, 29.2, 29.1, 28.9, 24.8, 20.9; IR (neat): 3004, 2931, 2858, 2225, 1731, 1585, 1469, 1434, 1373, 1265, 1230, 1110, 1072, 1026, 960 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{22}\text{H}_{29}\text{NO}_3\text{Na}$: 410.1943 $[\text{M}+\text{Na}]^+$; found: 410.1940.

General procedure for the synthesis of 6-oxo-4-substituted aryl-2-sulfanyl-1,6-dihydropyrimidine-5-carbonitrile (14a-14i)

A mixture with equimolar amounts of various aromatic aldehydes (**11a-11i**) (10 mmol), ethyl cyanoacetate (**12**) (10 mmol), thiourea (**13**) (10 mmol) and potassium carbonate (10 mmol) in absolute ethanol (25 mL) was gently heated under reflux temperature for 5 h. The completion of the reaction was monitored by TLC, upon completion a solid precipitated. The

reaction mixture was poured in ice water and stirred until clear solution was formed. Then the mixture was neutralized with glacial acetic acid to precipitate out the product, which was gently washed with water. The product was isolated and recrystallized from ethanol. The spectroscopic and analytical data of all the synthesized compounds were in good agreement with those reported in the literature.²⁷

General procedure for the synthesis of (E)-2-(2-Cyano-10-(3-methoxyphenyl)dec-2-enylthio)-6-oxo-4-phenyl-1,6-dihydropyrimidine-5-carbonitrile (15a-15t)

Potassium carbonate (1 mmol) was added to a well stirred solution of aryl-2-sulfanyl-1,6-dihydropyrimidine-5-carbonitrile (**14a-14i**) (1 mmol) in dry acetone (15 mL) at room temperature under nitrogen atmosphere and reaction mixture was stirred for 15 min at that temperature. Then Baylis-Hillman acetate (**10b-10g**) (1 mmol) in acetone (5 mL) was added slowly and the mixture was allowed to stir at reflux temperature for 2 h. Completion of the reaction was monitored by TLC. Following the completion of reaction, solvent was removed under reduced pressure, residue was diluted with water (15 mL) and extracted with ethyl acetate (2x25 mL). The combined organic layers were dried over sodium sulphate, filtered, concentrated under vacuum to give the crude product that was purified by silica gel column chromatography using ethyl acetate:hexane (3:7) as eluent, providing corresponding (E)-2-(2-Cyano-10-(3-methoxyphenyl)dec-2-enylthio)-6-oxo-4-phenyl-1,6-dihydropyrimidine-5-carbonitrile products (**15a-15t**) in good yields.

(E)-2-(2-Cyano-10-(3-methoxyphenyl)dec-2-enylthio)-6-oxo-4-phenyl-1,6-dihydropyrimidine-5-carbonitrile (15a): Sticky colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.94 (d, $J = 7.0$ Hz, 2H), 7.55-7.40 (m, 3H), 7.18 (t, $J = 8.3$ Hz, 1H), 6.79-6.69 (m, 3H), 6.47-6.39 (m, 1H), 4.10-4.00 (m, 2H), 3.78 (s, 3H), 2.55 (t, $J = 7.3$ Hz, 2H), 2.31-2.10 (m, 2H), 1.62-1.49 (m, 2H), 1.34-1.10 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 168.4, 159.5, 152.3, 144.3, 135.1, 131.9, 129.1, 128.9, 128.58, 128.5, 120.8, 116.3, 114.28, 114.2, 110.8, 110.7, 109.7, 102.3, 55.1, 35.9, 33.6, 31.5, 31.1, 29.0, 28.9, 28.8, 28.0; IR (neat): 2927, 2854, 2217, 1689, 1662, 1581, 1535, 1469, 1438, 1377, 1296, 1257, 1153, 1041, 1002 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{29}\text{H}_{31}\text{N}_4\text{O}_2\text{S}$: 499.2168 $[\text{M}+\text{H}]^+$; found: 499.2187.

(E)-2-(2-Cyano-10-(3-methoxyphenyl)dec-2-enylthio)-4-(4-methoxyphenyl)-6-oxo-1,6-dihydropyrimidine-5-carbonitrile (15b): Sticky light yellow liquid; ^1H NMR (300 MHz, CDCl_3): δ 8.01 (d, $J = 8.7$ Hz, 2H), 7.17 (t, $J = 7.7$ Hz, 1H), 6.93 (d, $J = 8.7$ Hz, 2H), 6.78-6.69 (m, 3H), 6.45 (t, $J = 7.6$ Hz, 1H), 4.13-4.03 (m, 2H), 3.84 (s, 3H), 3.78 (s, 3H), 2.54 (t, $J = 7.6$ Hz, 2H), 2.34-2.15 (m, 2H), 1.63-1.50 (m, 2H), 1.35-1.15 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 167.2, 164.3, 162.8, 159.5, 152.3, 144.3, 131.1, 129.1, 127.0, 120.7, 116.2, 114.2, 114.1, 113.9, 110.79, 110.7, 109.6, 91.6, 55.4, 55.0, 35.8, 33.5, 31.5, 31.1, 29.1, 29.0, 28.8, 28.0; IR (neat): 3432, 2927, 2854, 2217, 1650, 1604, 1539, 1508, 1469, 1377, 1303, 1261, 1176, 1153, 1029, 999, 914, 840, 783 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{30}\text{H}_{33}\text{N}_4\text{O}_3\text{S}$: 529.2273 $[\text{M}+\text{H}]^+$; found: 529.2282.

(E)-2-(2-Cyano-10-(3-methoxyphenyl)dec-2-enylthio)-4-(4-fluorophenyl)-6-oxo-1,6-dihydropyrimidine-5-carbonitrile

(15c): Sticky colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.85 (t, $J = 8.4$ Hz, 2H), 7.17 (t, $J = 8.0$ Hz, 1H), 7.02 (t, $J = 8.4$ Hz, 2H), 6.78-6.69 (m, 3H), 6.37 (t, $J = 7.4$ Hz, 1H), 5.37 (brs, 1H), 3.94 (s, 2H), 3.78 (s, 3H), 2.55 (t, $J = 7.6$ Hz, 2H), 2.29-2.01 (m, 2H), 1.65-1.51 (m, 2H), 1.37-1.12 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 167.6, 166.7, 166.2, 162.9, 159.5, 152.0, 144.3, 131.3, 131.1, 131.0, 129.1, 120.8, 116.9, 116.4, 115.7, 115.4, 114.1, 110.7, 110.0, 91.8, 55.0, 35.8, 33.6, 31.5, 31.1, 29.1, 29.0, 28.9, 28.1; IR (neat): 3440, 2927, 2854, 2217, 1662, 1604, 1539, 1512, 1485, 1469, 1380, 1303, 1257, 1242, 1157, 1103, 1045, 1006, 867, 837, 783 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{29}\text{H}_{30}\text{FN}_4\text{O}_2\text{S}$: 517.2074 $[\text{M}+\text{H}]^+$; found: 517.2074.

(E)-4-(4-Bromophenyl)-2-(2-cyano-10-(3-methoxyphenyl)dec-2-enylthio)-6-oxo-1,6-dihydropyrimidine-5-carbonitrile

(15d): Sticky light yellow liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.66 (d, $J = 8.3$ Hz, 2H), 7.45 (d, $J = 8.3$ Hz, 2H), 7.17 (t, $J = 7.6$ Hz, 1H), 6.78-6.68 (m, 3H), 6.40-6.30 (m, 1H), 4.90 (brs, 1H), 3.98-3.78 (m, 2H), 3.77 (s, 3H), 2.54 (t, $J = 7.6$ Hz, 2H), 2.28-2.06 (m, 2H), 1.62-1.49 (m, 2H), 1.30-1.08 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 168.2, 166.7, 166.6, 159.4, 151.9, 144.3, 134.0, 131.6, 130.1, 130.0, 129.0, 120.7, 117.1, 116.4, 114.1, 110.7, 110.0, 91.7, 55.0, 35.8, 33.8, 31.5, 31.1, 29.0, 28.9, 28.1; IR (neat): 3429, 2927, 2854, 2217, 1666, 1585, 1539, 1469, 1396, 1377, 1307, 1299, 1257, 1153, 1114, 1072, 1045, 1006, 860, 837, 802, 790, 694 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{29}\text{H}_{30}\text{BrN}_4\text{O}_2\text{S}$: 577.1273 $[\text{M}+\text{H}]^+$; found: 577.1266.

(E)-4-(3-Bromophenyl)-2-(2-cyano-10-(3-methoxyphenyl)dec-2-enylthio)-6-oxo-1,6-dihydropyrimidine-5-carbonitrile

(15e): Sticky light yellow liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.83 (s, 1H), 7.72 (d, $J = 7.8$ Hz, 1H), 7.51 (d, $J = 7.8$ Hz, 1H), 7.22-7.13 (m, 2H), 6.77-6.68 (m, 3H), 6.37 (t, $J = 7.6$ Hz, 1H), 4.45 (brs, 1H), 3.89 (s, 2H), 3.77 (s, 3H), 2.54 (t, $J = 7.6$ Hz, 2H), 2.28-2.07 (m, 2H), 1.60-1.49 (m, 2H), 1.30-1.12 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 168.9, 168.6, 166.2, 159.4, 151.9, 144.3, 137.2, 134.2, 131.4, 129.9, 129.1, 127.2, 122.5, 120.8, 116.9, 116.5, 114.1, 110.7, 110.2, 91.9, 55.1, 35.9, 33.8, 31.6, 31.2, 29.1, 29.0, 28.9, 28.1; IR (neat): 3421, 2927, 2854, 2217, 1677, 1539, 1485, 1465, 1377, 1303, 1257, 1211, 1153, 1076, 1045, 1010, 883, 783, 694 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{29}\text{H}_{30}\text{BrN}_4\text{O}_2\text{S}$: 577.1273 $[\text{M}+\text{H}]^+$; found: 577.1244.

(E)-4-(4-Chlorophenyl)-2-((2-cyano-10-(3-methoxyphenyl)dec-2-en-1-yl)thio)-6-oxo-1,6-dihydropyrimidine-5-carbonitrile

(15f): Sticky colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.81 (d, $J = 7.6$ Hz, 2H), 7.35 (d, $J = 7.6$ Hz, 2H), 7.17 (t, $J = 7.7$ Hz, 1H), 6.78-6.67 (m, 3H), 6.37 (t, $J = 7.7$ Hz, 1H), 4.99 (brs, 1H), 3.97 (s, 2H), 3.78 (s, 3H), 2.55 (t, $J = 7.6$ Hz, 2H), 2.30-2.08 (m, 2H), 1.63-1.49 (m, 2H), 1.34-1.12 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 166.7, 159.4, 152.5, 144.3, 138.1, 133.4, 130.1, 129.1, 128.7, 120.8, 116.3, 114.1, 110.7, 109.8, 55.0, 35.8, 33.7, 31.5, 31.1, 29.0, 28.9, 28.1; IR (neat): 3429, 2927, 2854, 2217, 1658, 1593, 1539, 1488, 1400, 1377, 1307, 1292, 1257, 1153, 1091, 1045,

1002, 840, 783 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{29}\text{H}_{30}\text{ClN}_4\text{O}_2\text{S}$: 533.1778 $[\text{M}+\text{H}]^+$; found: 533.1807.

(E)-4-(3-Chlorophenyl)-2-(2-cyano-10-(3-methoxyphenyl)dec-2-enylthio)-6-oxo-1,6-dihydropyrimidine-5-carbonitrile

(15g): Sticky colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.69-7.61 (m, 2H), 7.35-7.28 (m, 1H), 7.25-7.11 (m, 2H), 6.75-6.65 (m, 3H), 6.34 (t, $J = 7.6$ Hz, 1H), 5.46 (brs, 1H), 3.87 (s, 2H), 3.75 (s, 3H), 2.51 (t, $J = 7.4$ Hz, 2H), 2.25-2.01 (m, 2H), 1.59-1.47 (m, 2H), 1.31-1.06 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 168.8, 168.6, 166.2, 159.4, 151.9, 144.3, 137.0, 134.3, 131.2, 129.6, 129.0, 128.5, 126.7, 120.7, 116.9, 116.5, 114.1, 110.7, 110.1, 91.9, 55.0, 35.8, 33.7, 31.5, 31.2, 29.0, 28.9, 28.1; IR (neat): 3467, 2927, 2854, 2217, 1677, 1581, 1539, 1485, 1469, 1377, 1303, 1257, 1211, 1153, 1045, 1010, 891, 783 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{29}\text{H}_{30}\text{ClN}_4\text{O}_2\text{S}$: 533.1778 $[\text{M}+\text{H}]^+$; found: 533.1773.

(E)-2-(2-Cyano-10-(3-methoxyphenyl)dec-2-enylthio)-6-oxo-4-(thiophen-2-yl)-1,6-dihydropyrimidine-5-carbonitrile

(15h): Sticky colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 8.28 (d, $J = 3.6$ Hz, 1H), 7.57 (d, $J = 4.6$ Hz, 1H), 7.17 (t, $J = 7.6$ Hz, 1H), 7.09 (t, $J = 4.6$ Hz, 3.6 Hz, 1H), 6.77-6.66 (m, 3H), 6.58 (t, $J = 7.4$ Hz, 1H), 5.85 (brs, 1H), 4.07-3.93 (m, 2H), 3.78 (s, 3H), 2.53 (t, $J = 7.8$ Hz, 2H), 2.36-2.21 (m, 2H), 1.62-1.49 (m, 2H), 1.40-1.16 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 165.3, 165.1, 159.5, 159.4, 152.0, 144.3, 139.5, 133.6, 132.2, 129.3, 129.1, 120.7, 116.3, 116.2, 114.1, 110.7, 109.8, 88.1, 55.0, 35.8, 33.3, 31.5, 31.1, 29.1, 29.0, 28.9, 28.1; IR (neat): 3429, 2927, 2854, 2217, 1666, 1581, 1546, 1527, 1469, 1415, 1388, 1346, 1288, 1257, 1149, 1114, 1045, 983, 864, 779, 725 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{27}\text{H}_{29}\text{N}_4\text{O}_2\text{S}_2$: 505.1732 $[\text{M}+\text{H}]^+$; found: 505.1767.

(E)-2-(2-Cyano-10-(3-methoxyphenyl)dec-2-enylthio)-4-(3,4-dichlorophenyl)-6-oxo-1,6-dihydropyrimidine-5-carbonitrile

(15i): Sticky light yellow liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.88-7.85 (m, 1H), 7.70 (d, $J = 8.4$ Hz, 1H), 7.43 (d, $J = 8.4$ Hz, 1H), 7.17 (t, $J = 7.7$ Hz, 1H), 6.78-6.68 (m, 3H), 6.39 (t, $J = 7.4$ Hz, 1H), 4.07 (brs, 1H), 3.95 (s, 2H), 3.77 (s, 3H), 2.54 (t, $J = 7.4$ Hz, 2H), 2.31-2.11 (m, 2H), 1.63-1.51 (m, 2H), 1.36-1.15 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 168.3, 165.2, 159.4, 152.1, 144.2, 135.9, 134.7, 132.8, 130.7, 130.3, 129.0, 127.6, 120.6, 116.4, 116.3, 114.0, 110.6, 109.9, 92.1, 55.0, 35.8, 33.7, 31.5, 31.1, 28.9, 28.8, 28.0 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{29}\text{H}_{29}\text{Cl}_2\text{N}_4\text{O}_2\text{S}$: 567.1467 $[\text{M}+\text{H}]^+$; found: 567.1391.

(E)-2-(2-Cyano-10-(3-propoxyphenyl)dec-2-enylthio)-6-oxo-4-phenyl-1,6-dihydropyrimidine-5-carbonitrile

(15j): Sticky colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.77-7.67 (m, 2H), 7.39-7.32 (m, 1H), 7.31-7.22 (m, 2H), 7.14 (t, $J = 7.7$ Hz, 1H), 6.75-6.65 (m, 3H), 6.32 (t, $J = 7.6$ Hz, 1H), 5.11 (brs, 1H), 3.93-3.91 (m, 4H), 2.52 (t, $J = 7.7$ Hz, 2H), 2.22-2.04 (m, 2H), 1.84-1.71 (sex, $J = 7.1$, 6.8 Hz, 2H), 1.60-1.47 (m, 2H), 1.27-1.07 (m, 8H), 1.01 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.7, 169.1, 167.8, 159.1, 151.7, 144.2, 135.5, 131.1, 129.0, 128.6, 128.3, 120.6, 118.0, 116.6, 114.7, 111.3, 110.2, 91.3, 69.3, 35.9, 33.6, 31.5, 31.2, 29.0, 28.9, 28.1, 22.6, 10.5; IR (neat): 2927, 2854, 2217, 1670, 1581, 1539, 1473, 1438, 1315, 1296, 1257, 1157, 1002, 860, 767, 694, 675 cm^{-1} ;

HRMS-ESI (m/z): Calcd for $C_{31}H_{35}N_4O_2S$: 527.2481 $[M+H]^+$; found: 527.2482.

(E)-2-(10-(3-Butoxyphenyl)-2-cyanodec-2-enylthio)-6-oxo-4-phenyl-1,6-dihydropyrimidine-5-carbonitrile (15k): Sticky colorless liquid; 1H NMR (300 MHz, $CDCl_3$): δ 7.81 (d, $J = 7.4$ Hz, 2H), 7.41 (d, $J = 7.3$ Hz, 1H), 7.34 (t, $J = 7.4, 7.3$ Hz, 2H), 7.16 (t, $J = 7.3$ Hz, 1H), 6.76-6.67 (m, 3H), 6.36 (t, $J = 7.6$ Hz, 1H), 5.49 (brs, 1H), 3.98-3.88 (m, 4H), 2.53 (t, $J = 7.7$ Hz, 2H), 2.25-2.07 (m, 2H), 1.80-1.69 (m, 2H), 1.61-1.42 (m, 4H), 1.26-1.10 (m, 8H), 0.96 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (75 MHz, $CDCl_3$): δ 168.2, 168.0, 167.9, 159.0, 151.9, 144.2, 135.3, 131.3, 129.0, 128.7, 128.3, 120.5, 116.5, 114.8, 114.7, 111.3, 110.0, 92.0, 67.4, 35.8, 33.6, 31.5, 31.3, 31.2, 29.0, 28.98, 28.9, 28.0, 19.2, 13.8; IR (neat): 2927, 2854, 2217, 1666, 1562, 1535, 1481, 1469, 1299, 1253, 1157, 1006 cm^{-1} ; HRMS-ESI (m/z): Calcd for $C_{32}H_{37}N_4O_2S$: 541.2637 $[M+H]^+$; found: 541.2665.

(E)-2-(2-Cyano-10-(3-(pentyloxy)phenyl)dec-2-enylthio)-6-oxo-4-phenyl-1,6-dihydropyrimidine-5-carbonitrile (15l): Sticky colorless liquid; 1H NMR (300 MHz, $CDCl_3$): δ 7.79 (d, $J = 7.7$ Hz, 2H), 7.40 (d, $J = 7.0$ Hz, 1H), 7.32 (t, $J = 7.7, 7.0$ Hz, 2H), 7.15 (t, $J = 7.6$ Hz, 1H), 6.75-6.67 (m, 3H), 6.35 (t, $J = 7.6$ Hz, 1H), 4.15 (brs, 1H), 3.97-3.87 (m, 4H), 2.53 (t, $J = 7.6$ Hz, 2H), 2.25-2.04 (m, 2H), 1.81-1.72 (quin, $J = 6.8$ Hz, 2H), 1.61-1.50 (m, 2H), 1.48-1.33 (m, 4H), 1.29-1.12 (m, 8H), 0.91 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (75 MHz, $CDCl_3$): δ 169.1, 168.1, 167.9, 159.1, 151.7, 144.2, 135.6, 135.5, 131.1, 129.0, 128.6, 128.3, 120.5, 116.6, 114.7, 111.3, 110.2, 91.4, 67.7, 35.9, 33.6, 31.4, 31.2, 29.0, 28.9, 28.17, 28.1, 22.4, 19.4, 13.9; IR (neat): 3371, 2927, 2854, 2217, 1674, 1581, 1535, 1473, 1438, 1380, 1315, 1299, 1257, 1157, 1018, 771, 694 cm^{-1} ; HRMS-ESI (m/z): Calcd for $C_{33}H_{39}N_4O_2S$: 555.2794 $[M+H]^+$; found: 555.2793.

(E)-2-(2-Cyano-10-(3-(hexyloxy)phenyl)dec-2-enylthio)-6-oxo-4-phenyl-1,6-dihydropyrimidine-5-carbonitrile (15m): Sticky colorless liquid; 1H NMR (300 MHz, $CDCl_3$): δ 7.90 (d, $J = 7.4$ Hz, 2H), 7.52-7.32 (m, 3H), 7.16 (t, $J = 7.4$ Hz, 1H), 6.76-6.68 (m, 3H), 6.39 (t, $J = 7.6$ Hz, 1H), 4.06-3.98 (m, 2H), 3.93 (t, $J = 6.4$ Hz, 2H), 3.54 (brs, 1H), 2.54 (t, $J = 7.6$ Hz, 2H), 2.30-2.10 (m, 2H), 1.81-1.71 (quin, $J = 6.4$ Hz, 2H), 1.61-1.51 (m, 2H), 1.49-1.41 (m, 2H), 1.37-1.31 (m, 4H), 1.28-1.11 (m, 8H), 0.89 (t, $J = 6.5$ Hz, 3H); ^{13}C NMR (75 MHz, $CDCl_3$): δ 168.1, 167.4, 167.0, 159.0, 152.0, 144.2, 135.2, 131.5, 129.0, 128.7, 128.4, 120.5, 116.5, 116.4, 114.7, 111.3, 109.9, 92.4, 67.7, 35.8, 33.6, 31.5, 31.3, 31.2, 29.2, 29.0, 28.9, 28.0, 25.6, 22.5, 13.9; IR (neat): 2927, 2854, 2217, 1666, 1581, 1539, 1473, 1438, 1377, 1296, 1253, 1157, 1006, 767, 694 cm^{-1} ; HRMS-ESI (m/z): Calcd for $C_{34}H_{41}N_4O_2S$: 569.2950 $[M+H]^+$; found: 569.2984.

(E)-Methyl 2-(9-cyano-10-(5-cyano-6-oxo-4-phenyl-1,6-dihydropyrimidin-2-ylthio)dec-8-enyl)-6-methoxybenzoate (15n): Sticky colorless liquid; 1H NMR (300 MHz, $CDCl_3$): δ 7.97 (d, $J = 7.1$ Hz, 2H), 7.59-7.44 (m, 3H), 7.26 (t, $J = 7.7$ Hz, 1H), 6.80 (d, $J = 7.7$ Hz, 1H), 6.76 (d, $J = 8.3$ Hz, 1H), 6.53-6.41 (m, 1H), 5.91 (brs, 1H), 4.15-4.04 (m, 2H), 3.89 (s, 3H), 3.80 (s, 3H), 2.51 (t, $J = 7.7$ Hz, 2H), 2.34-2.11 (m, 2H), 1.60-1.47 (m, 2H), 1.33-1.13 (m, 8H); ^{13}C NMR (75 MHz, $CDCl_3$): δ 169.0, 168.9, 168.4, 164.4, 156.2, 152.5, 141.0, 134.8, 132.1,

130.2, 129.0, 128.9, 128.6, 128.5, 121.4, 116.1, 109.6, 108.4, 101.0, 55.8, 52.1, 33.6, 33.3, 31.5, 30.9, 29.1, 28.9, 28.7, 27.9; IR (neat): 2927, 2854, 2221, 1728, 1693, 1662, 1581, 1535, 1469, 1434, 1377, 1265, 1110, 1072, 1002, 767, 698 cm^{-1} ; HRMS-ESI (m/z): Calcd for $C_{31}H_{33}N_4O_4S$: 557.2223 $[M+H]^+$; found: 557.2258.

(E)-Methyl 2-(9-cyano-10-(5-cyano-4-(4-methoxyphenyl)-6-oxo-1,6-dihydropyrimidin-2-ylthio)dec-8-enyl)-6-methoxybenzoate (15o): Colorless liquid; 1H NMR (300 MHz, $CDCl_3$): δ 8.03 (d, $J = 8.7$ Hz, 2H), 7.25 (t, $J = 8.1$ Hz, 1H), 6.95 (d, $J = 8.7$ Hz, 2H), 6.79 (d, $J = 7.6$ Hz, 1H), 6.75 (d, $J = 8.1$ Hz, 1H), 6.46 (t, $J = 7.7$ Hz, 1H), 4.10-4.03 (m, 2H), 3.89 (s, 3H), 3.85 (s, 3H), 3.79 (s, 3H), 2.50 (t, $J = 7.6$ Hz, 2H), 2.34-2.08 (m, 2H), 1.61-1.46 (m, 2H), 1.33-1.13 (m, 8H); ^{13}C NMR (75 MHz, $CDCl_3$): δ 168.9, 167.2, 163.9, 162.9, 156.1, 152.3, 141.0, 131.1, 130.2, 126.9, 123.3, 121.4, 116.1, 115.8, 113.9, 109.6, 108.3, 91.8, 55.8, 55.4, 52.1, 33.4, 33.3, 31.5, 30.8, 29.1, 28.9, 28.7, 28.0; IR (neat): 2931, 2854, 2217, 1728, 1689, 1654, 1604, 1539, 1469, 1377, 1307, 1261, 1180, 1110, 1072, 1026, 999, 840, 783 cm^{-1} ; HRMS-ESI (m/z): Calcd for $C_{32}H_{35}N_4O_5S$: 587.2328 $[M+H]^+$; found: 587.2322.

(E)-Methyl 2-(9-cyano-10-(5-cyano-4-(4-fluorophenyl)-6-oxo-1,6-dihydropyrimidin-2-ylthio)dec-8-enyl)-6-methoxybenzoate (15p): Sticky colorless liquid; 1H NMR (300 MHz, $CDCl_3$): δ 7.87-7.78 (m, 2H), 7.22 (t, $J = 7.8$ Hz, 1H), 7.01 (t, $J = 8.4$ Hz, 2H), 6.76 (d, $J = 7.7$ Hz, 1H), 6.72 (d, $J = 8.3$ Hz, 1H), 6.34 (t, $J = 7.7$ Hz, 1H), 5.57 (brs, 1H), 3.99-3.89 (m, 2H), 3.86 (s, 3H), 3.77 (s, 3H), 2.48 (t, $J = 7.7$ Hz, 2H), 2.25-2.06 (m, 2H), 1.55-1.43 (m, 2H), 1.30-1.08 (m, 8H); ^{13}C NMR (75 MHz, $CDCl_3$): δ 169.0, 168.2, 167.9, 166.6, 162.9, 156.2, 151.9, 141.0, 131.1, 131.0, 130.2, 123.3, 121.4, 117.0, 116.4, 115.6, 115.3, 110.0, 108.3, 91.6, 55.8, 52.1, 33.6, 33.3, 31.4, 30.8, 29.1, 28.9, 28.7, 28.0; IR (neat): 3452, 2931, 2854, 2217, 1731, 1689, 1600, 1581, 1539, 1508, 1469, 1373, 1307, 1265, 1242, 1141, 1087, 1072, 1060, 1006, 914, 848 cm^{-1} ; HRMS-ESI (m/z): Calcd for $C_{31}H_{32}FN_4O_4S$: 575.2128 $[M+H]^+$; found: 575.2136.

(E)-Methyl 2-(10-(4-(4-bromophenyl)-5-cyano-6-oxo-1,6-dihydropyrimidin-2-ylthio)-9-cyanodec-8-enyl)-6-methoxybenzoate (15q): Sticky light yellow liquid; 1H NMR (300 MHz, $CDCl_3$): δ 7.79 (d, $J = 8.3$ Hz, 2H), 7.57 (d, $J = 8.3$ Hz, 2H), 7.25 (t, $J = 7.7$ Hz, 1H), 6.79 (d, $J = 7.7$ Hz, 1H), 6.75 (d, $J = 8.3$ Hz, 1H), 6.40 (t, $J = 7.6$ Hz, 1H), 4.07-3.99 (m, 2H), 3.89 (s, 3H), 3.80 (s, 3H), 2.51 (t, $J = 7.6$ Hz, 2H), 2.32-2.11 (m, 2H), 1.60-1.47 (m, 2H), 1.33-1.15 (m, 8H); ^{13}C NMR (75 MHz, $CDCl_3$): δ 169.0, 167.2, 167.0, 165.2, 156.2, 152.4, 141.0, 133.7, 131.8, 130.3, 130.2, 126.9, 123.3, 121.4, 116.2, 115.5, 109.7, 108.3, 93.1, 55.8, 52.1, 33.6, 33.3, 31.5, 31.5, 29.1, 28.9, 28.7, 28.0; IR (neat): 3436, 2927, 2854, 2217, 1728, 1693, 1662, 1585, 1535, 1469, 1299, 1265, 1110, 1072, 1006, 837, 771 cm^{-1} ; HRMS-ESI (m/z): Calcd for $C_{31}H_{32}BrN_4O_4S$: 635.1328 $[M+H]^+$; found: 635.1330.

(E)-Methyl 2-(10-(4-(4-chlorophenyl)-5-cyano-6-oxo-1,6-dihydropyrimidin-2-ylthio)-9-cyanodec-8-enyl)-6-methoxybenzoate (15r): Sticky light yellow liquid; 1H NMR (300 MHz, $CDCl_3$): δ 7.86 (d, $J = 8.4$ Hz, 2H), 7.39 (d, $J = 8.4$

Hz, 2H), 7.25 (t, $J = 7.8$ Hz, 1H), 6.79 (d, $J = 7.8$ Hz, 1H), 6.75 (d, $J = 8.3$ Hz, 1H), 6.40 (t, $J = 7.6$ Hz, 1H), 4.07-3.99 (m, 2H), 3.89 (s, 3H), 3.79 (s, 3H), 2.51 (t, $J = 7.6$ Hz, 2H), 2.32-2.12 (m, 2H), 1.59-1.47 (m, 2H), 1.33-1.15 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.0, 166.9, 166.1, 165.4, 156.2, 152.4, 141.1, 138.3, 133.3, 130.3, 128.9, 123.3, 121.4, 118.6, 116.3, 115.7, 109.8, 108.4, 93.1, 55.8, 52.2, 33.7, 33.3, 31.6, 30.9, 29.2, 28.9, 28.8, 28.1; IR (neat): 3440, 2931, 2854, 2217, 1728, 1693, 1662, 1581, 1539, 1469, 1377, 1307, 1265, 1110, 1091, 1083, 1002, 840, 786 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{31}\text{H}_{32}\text{ClN}_4\text{O}_4\text{S}$: 591.1833 [$\text{M}+\text{H}$] $^+$; found: 591.1828.

(E)-methyl 2-(10-(4-(3-chlorophenyl)-5-cyano-6-oxo-1,6-dihydropyrimidin-2-ylthio)-9-cyanodec-8-enyl)-6-methoxybenzoate (15s): Sticky colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.71-7.63 (m, 2H), 7.39-7.31 (m, 1H), 7.28-7.18 (m, 2H), 6.78 (d, $J = 7.8$ Hz, 1H), 6.74 (d, $J = 8.3$ Hz, 1H), 6.35 (t, $J = 7.7$ Hz, 1H), 5.59 (brs, 1H), 3.95-3.83 (m, 5H), 3.78 (s, 3H), 2.49 (t, $J = 7.6$ Hz, 2H), 2.26-2.07 (m, 2H), 1.57-1.44 (m, 2H), 1.27-1.10 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.5, 169.0, 167.5, 166.2, 156.1, 151.7, 143.1, 141.0, 134.3, 130.2, 129.7, 128.4, 126.6, 123.2, 121.4, 120.9, 116.5, 115.8, 110.3, 108.3, 91.5, 55.8, 52.1, 33.8, 33.3, 31.5, 30.9, 29.1, 28.9, 28.8, 28.0; IR (neat): 3429, 2927, 2854, 2217, 1728, 1647, 1581, 1539, 1469, 1380, 1303, 1265, 1207, 1161, 1110, 1072, 1010, 891, 790 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{31}\text{H}_{32}\text{ClN}_4\text{O}_4\text{S}$: 591.1833 [$\text{M}+\text{H}$] $^+$; found: 591.1835

(E)-Methyl 2-(9-cyano-10-(5-cyano-4-(3,4-dichlorophenyl)-6-oxo-1,6-dihydropyrimidin-2-ylthio)dec-8-enyl)-6-methoxybenzoate (15t): Sticky colorless liquid; ^1H NMR (300 MHz, CDCl_3): δ 7.83 (s, 1H), 7.66 (d, $J = 8.3$ Hz, 1H), 7.40 (d, $J = 8.3$ Hz, 1H), 7.24 (t, $J = 8.1$ Hz, 1H), 6.79 (d, $J = 7.7$ Hz, 1H), 6.74 (d, $J = 8.1$ Hz, 1H), 6.37 (t, $J = 7.6$ Hz, 1H), 4.27 (brs, 1H), 3.97-3.86 (m, 5H), 3.79 (s, 3H), 2.50 (t, $J = 7.6$ Hz, 2H), 2.30-2.08 (m, 2H), 1.59-1.46 (m, 2H), 1.33-1.11 (m, 8H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.0, 168.5, 168.0, 164.4, 155.5, 150.3, 140.3, 136.1, 133.9, 131.8, 129.8, 129.7, 127.3, 122.8, 120.8, 117.2, 116.1, 110.5, 107.8, 90.8, 55.2, 51.4, 32.8, 32.6, 30.8, 30.3, 28.5, 28.3, 28.1, 27.5 cm^{-1} ; HRMS-ESI (m/z): Calcd for $\text{C}_{31}\text{H}_{31}\text{Cl}_2\text{N}_4\text{O}_4\text{S}$: 625.1443 [$\text{M}+\text{H}$] $^+$; found: 625.1446.

CONCLUSION

We have described a convenient and efficient protocol for the synthesis of novel Baylis-Hillman adducts and their acetates in fine yields under solvent-free conditions from inexpensive CNSL constituents and, evaluated for anticancer activity. The novelty of this protocol is mainly related to the use of CNSL components as starting materials for Baylis-Hillman adducts. Compounds **5b** and **5h** bearing methoxy functionality in the phenyl ring showed promising activity against MCF-7. Furthermore, we have described the synthesis, biological activity and structure-activity relationship of a series of twenty novel phenolic based pyrimidine derivatives. Baylis-Hillman adducts derived from CNSL renewable natural resources were employed as key precursors in the synthesis of these pyrimidine derivatives. Compounds **15i**, **15j** and **15l** showed promising activity against HEP-G2 with the GI_{50} value of 6.17, 2.87 and

4.56 μM respectively, whereas compounds **15j**, **15k**, **15l** and **15m** showed promising activity against MCF-7 with the GI_{50} value of 8.51, 8.64, 8.66 and 5.02 μM respectively. Compound **15m** showed high activity against K562 cell line with the GI_{50} value of 0.25 μM being similar to standard control. These results illustrated that alkyl substitutions on the aromatic ring are important for maintaining the antitumor activity. Our structure activity relationship (SAR) study around these scaffolds suggests that new antitumor agents with an improved biological profile can be prepared for developing new clinical candidates.

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SUPPLEMENTARY INFORMATION

General experimental procedures, characterization details of **2-3**, **7-8**, and ^1H and ^{13}C NMR spectra of all synthesized compounds.

REFERENCES AND NOTES

- (a) J.R. Benson, I. Jatoi. The global breast cancer burden. *Future Oncol.* **2012**, *8*, 697-702; (b) M. Kavallaris. Microtubules and resistance to tubulin-binding agents. *Nat. Rev. Cancer*, **2010**, *10*, 194-204.
- E.T. Hawk, E.B. Habermann, J.G. Ford, J.A. Wenzel, J.R. Brahma, M.S. Chen, L.A. Jones, T.C. Hurd, L.M. Rogers, L.H. Nguyen, J.S. Ahluwalia, M. Fouad, S.M. Vickers. You have free access to this content Five National Cancer Institute-designated cancer centers' data collection on racial/ethnic minority participation in therapeutic trials: A current view and opportunities for improvement. *Cancer*, **2014**, *1*, 1113-1121
- H. Varmus. The new era in cancer research. *Science*, **2006**, *312*, 1162-1165.
- D.J. Newman, G.M. Cragg. Natural products as sources of new drugs over the 30 years from 1981 to 2010. *J. Nat. Prod.* **2012**, *75*, 311-335.
- C. Voirin, S. Caillol, N.V. Sadavarte, B.V. Tawade, B. Boutevin, P.P. Wadgaonkar. Functionalization of cardanol: towards biobased polymers and additives. *Polym. Chem.* **2014**, *5*, 3142-3162.
- V.S. Balachandran, S.R. Jadhav, P.K. Vemula, G. John. Recent advances in cardanol chemistry in a nutshell: from a nut to nanomaterials. *Chem. Soc. Rev.* **2013**, *42*, 427-438.
- E.B. Mubofu. From cashew nut shell wastes to high value chemicals. *Pure Appl. Chem.* **2016**, *88*, 17-27.
- J.H.P. Tyman. Synthetic and natural phenols, Elsevier, Amsterdam, **1996**; (b) O.A. Attanasi, P. Filippone. *Chim. Ind. (Milano)* **2003**, *85*, 11.
- P.K. Phani, R. Paramashivappa, P.J. Vithayathil, P.V. Subba Rao, S. Rao. Process for isolation of cardanol from technical cashew (*Anacardium occidentale* L.) nut shell liquid. *J. Agric. Food Chem.* **2002**, *50*, 4705-4708.
- R. Paramashivappa, P.P. Kumar, P.J. Vithayathil, A.S. Rao. Novel method for isolation of major phenolic constituents from cashew (*Anacardium occidentale* L.) nut shell liquid. *J. Agric. Food Chem.* **2001**, *49*, 2548-2551.
- (a) M.C. Lubi, E.B. Thachill. Cashew nut shell liquid (CNSL) a versatile monomer for polymer synthesis. *Des. Monomers Polym.* **2000**, *3*, 123-153; (b) E. Darroman, N. Durand, B. Boutevin, S. Caillol. New cardanol/sucrose epoxy blends for biobased coatings. *Prog. Org. Coat.* **2015**, *83*, 47-54.
- (a) I. Kubo, M. Ochi, P.C. Vieira, S. Komatsu. Antitumor agents from the cashew (*Anacardium occidentale*) apple juice. *J. Agric. Food*

- Chem.* **1993**, 41, 1012-1015; (b) G.K. Mahapatro. Insecticidal activity of cashew nut shell liquid against two lepidopteran pests. *Indian J. Entomol.* **2011**, 73, 121-124; (c) T.J. Ha, I. Kubo. Lipoxygenase inhibitory activity of anacardic acids. *J. Agric. Food Chem.* **2005**, 53, 4350-4354; (d) R. Amorati, G.F. Pedulli, L. Valgimigli, O.A. Attanasi, P. Filippone, C. Fiorucci, R. Saladino. Absolute rate constants for the reaction of peroxy radicals with cardanol derivatives. *J. Chem. Soc., Perkin Trans. 2* **2001**, 2142-2146; (e) M. Himejima, I. Kubo. Antibacterial agents from the cashew *Anacardium occidentale* (Anacardiaceae) nut shell oil. *J. Agric. Food Chem.* **1991**, 39, 418-421; (f) P.D. Adawadkar, M.A. El Sohly. Isolation, purification and antimicrobial activity of anacardic acids from Ginkgo biloba fruits. *Fitoterapia* **1981**, 52, 129-135; (g) M.S.C. Oliveira, S.M. Morais, D.V. Magalhães, W.P. Batista, I.G.P. Vieira, A.A. Craveiro, J.E.S.A. Menezes, A.F.U. Carvalho, G.P.G. Lima. Antioxidant, larvicidal and antiacetylcholinesterase activities of cashew nut shell liquid constituents. *Acta Trop.* **2010**, 117, 165-170.
13. O.A. Attanasi, S. Berretta, C. Fiani, P. Filippone, G. Mele, R. Saladino. Synthesis and reactions of nitro derivatives of hydrogenated cardanol. *Tetrahedron*, **2006**, 62, 6113-6120.
14. C. Voirin, S. Caillol, N.V. Sadavarte, B.V. Tawade, B. Boutevin, P.P. Wadgaonkar. Functionalization of cardanol: towards biobased polymers and additives. *Polym. Chem.* **2014**, 5, 3142-3162.
15. M.L. Santos, G.C. Magalhães. Utilisation of cashew nut shell liquid from *Anacardium occidentale* as starting material for organic synthesis: a novel route to lasiodiplodin from cardols. *J. Braz. Chem. Soc.* **1999**, 10, 13-20.
16. A.F. Pozharskii, A.T. Soldatenkov, A.R. Katritzky. In: Heterocycles and Health, in: Heterocycles in Life and Society, John Wiley & Sons, Chichester, UK, **1997**, p. 135-164.
17. (a) H.T. Abdel-Mohsen, F. A.F. Ragab, M.M. Ramla, H.I. El Diwani. Novel benzimidazole-pyrimidine conjugates as potent antitumor agents. *Eur. J. Med. Chem.* **2010**, 45, 2336-2344; (b) C. Srinivasulu, K. Rajshekhar, C. Balakumar, A.H. Girish, T. Neeta, N.P. Venkata. An insight on synthetic and medicinal aspects of pyrazolo[1,5-a]pyrimidine scaffold. *Eur. J. Med. Chem.* **2017**, 126, 298-352.
18. (a) H. Parveen, F. Hayat, A. Salahuddin, A. Azam. Synthesis, characterization and biological evaluation of novel 6-ferrocenyl-4-aryl-2-substituted pyrimidine derivatives. *Eur. J. Med. Chem.* **2010**, 45, 3497-3503; (b) S.A.F. Rostom, H.M.A. Ashour, H.A. Abd El Razik. Synthesis and biological evaluation of some novel polysubstituted pyrimidine derivatives as potential antimicrobial and anticancer agents. *Arch. Pharm.* **2009**, 342, 299-310; (c) O.A. Fathalla, S.M. Awad, M.S. Mohamed. Synthesis of new 2-thiouracil-5-sulphonamide derivatives with antibacterial and antifungal activity. *Arch. Pharm. Res.* **2005**, 28, 1205-1212; (d) D.J. Brown. Pyrimidines and their benzo derivatives. in: A. R. Katritzky, C. W. Rees (Eds.), *Comprehensive Heterocyclic Chemistry, the Structure, Reactions, Synthesis and Uses of Heterocyclic Compounds*, 3. Pergamon Press, Oxford, **1984**, pp. 57-155; (e) G. Vanessa, M. Sidnei, F.C.F. Alex, C.F. Darlene, C. Pio, P. Ernani. Antioxidant and antimicrobial properties of 2-(4,5-dihydro-1H-pyrazol-1-yl)-pyrimidine and 1-carboxamidino-1H-pyrazole derivatives. *J. Braz. Chem. Soc.* **2010**, 21, 1477-1483; (f) R. Bannella, S.P. Shrivastava. Synthesis and in vitro antimicrobial, anthelmintic and insecticidal activities study of 4(4'-bromophenyl)-6-substituted-aryl-1-acetyl pyrimidine-2-thiols. *E. -J. Chem.* **2010**, 7, 935-941; (g) F. Morales, A. Ramirez, A. Conejo-Garcia, C. Morata, J.A. Marchal, J.M. Campos. Anti-proliferative activity of 2,6-dichloro-9- or 7-(ethoxycarbonylmethyl)-9H- or 7H-purines against several human solid tumour cell lines. *Eur. J. Med. Chem.* **2014**, 76, 118-124; (h) B.S. Chhikara, B. Rathi. Organic molecular drugs designing for inhibition of cellular multiplication biocycle of virus for anti-HIV therapy development: A recent advances review. *Adv. Org. Chem. Lett.* **2015**, 2(1), 1-14; (i) D.J. Brown, R.F. Evans, The pyrimidines: suppl. 2. in: A. Weissberger, E.C. Taylor (Eds.), *The Chemistry of Heterocyclic Compounds*. John Wiley & Sons Inc., New Jersey, **1985**; (j) M. Johar, T. Manning, D.Y. Kunimoto, R. Kumara. Synthesis and in vitro antimycobacterial activity of 5-substituted pyrimidine nucleosides. *Bioorg. Med. Chem.* **2005**, 13, 6663-6671.
19. (a) X. Li, J. Yang, X. Chen, J. Liu, H. Li, J. Zheng, Y. He, Z. Chen, S. Huang. A report of early cytogenetic response to imatinib in two patients with chronic myeloid leukemia at accelerated phase and carrying the e19a2 BCR-ABL transcript. *Cancer Genet. Cytogenet.* **2007**, 176, 166-168; (b) I. El Hajj Dib, M. Gallet, R. Mentaverri, N. Sévenet, M. Brazier, S. Kamel. Imatinib mesylate (Gleevec) enhances mature osteoclast apoptosis and suppresses osteoclast bone resorbing activity. *Eur. J. Pharmacol.* **2006**, 551, 27-33.
20. For selected reviews on the Morita-Baylis-Hillman reaction see: (a) D. Basavaiah, B.S. Reddy, S.S. Badsara. Recent contributions from the Baylis-Hillman reaction to organic chemistry. *Chem. Rev.* **2010**, 110, 5447-5674; (b) T.Y. Liu, M. Xie, Y.C. Chen. Organocatalytic asymmetric transformations of modified Morita-Baylis-Hillman adducts. *Chem. Soc. Rev.* **2012**, 41, 4101-4112.
21. T. Narendar Reddy, C. Swetha, P. Ramesh, B. Sridhar, V. Jayathirtha Rao. Synthesis of phenylselenopyrans and lactones from allylic alcohols and acids via Baylis-Hillman reaction. *Chemistry Select*, **2017**, 2, 8402-8407 and references cited therein.
22. T. Narendar Reddy, M. Ravinder, R. Bikshapathi, P. Sujitha, C.G. Kumar, V. Jayathirtha Rao. Design, synthesis, and biological evaluation of 4-H pyran derivatives as antimicrobial and anticancer agents. *Med. Chem. Res.* **2017**, 26, 2832-2844 and references cited therein.
23. A.A.S. Naujorks, A.O. Da Silva, R.S. Lopes, S. De Albuquerque, A. Beatriz, M.R. Marques, D.P. De Lima. Novel naphthoquinone derivatives and evaluation of their trypanocidal and leishmanicidal activities. *Org. Biomol. Chem.* **2015**, 13, 428-437.
24. I. Kubo, S. Komatsu, M. Ochi. Molluscicides from the cashew *Anacardium occidentale* and their large-scale isolation. *J. Agr. Food Chem.* **1986**, 34, 970-973.
25. M.B. Graham, J.H.P. Tyman. Ozonization of phenols from *Anacardium occidentale* (cashew). *J. Am. Oil Chem. Soc.* **2002**, 79, 725-732.
26. R. Paramashivappa, P. Phanikumar, P.J. Vithayathil, A. Srinivasa Rao. Novel method for isolation of major phenolic constituents from cashew (*Anacardium occidentale* L.) nut shell liquid. *J. Agric. Food Chem.* **2001**, 49, 2548-2551.
27. L.-Y. Ma, L.-P. Pang, B. Wang, M. Zhang, B. Hu, D.-Q. Xue, K.-P. Shao, B.-L. Zhang, Y. Liu, E. Zhang, H.-M. Liu. Design and synthesis of novel 1,2,3-triazole-pyrimidine hybrids as potential anticancer agents. *Eur. J. Med. Chem.* **2014**, 86, 368-380.
28. (a) P. Skehan, R. Storeng, D. Scudiero, A. Monks, J. McMahon, D. Vistica, J.T. Warren, H. Bokesch, S. Kenney, M.R. Boyd. New colorimetric cytotoxicity assay for anticancer-drug screening. *J. Natl. Cancer Inst.* **1990**, 82, 1107-1112; (b) A. Monks, D. Scudiero, P. Skehan, R. Shoemaker, K. Pau, D. Vistica, H. Curtis, J. Langley, P. Cronise, A. Vaigro-Wolff, M. Gray-Goodrich, H. Campbell, J. Mayo, M. Boyd. Feasibility of a high-flux anticancer drug screen using a diverse panel of cultured human tumor cell lines. *J. Natl. Cancer Inst.* **1991**, 83, 757-766; (c) T. Mosmann. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J. Immunol. Methods*, **1983**, 65, 55-63.
29. (a) L.P.L. Logrado, C.O. Santos, L.A.S. Romeiro, A.M. Costa, J.R.O. Ferreira, C. Cavalcanti, O.M. de Moraes, L.V. Costa-Lotufo, C. Pessoa, M.L. dos Santos. Synthesis and cytotoxicity screening of substituted isobenzofuranones designed from anacardic acids. *Eur. J. Med. Chem.* **2010**, 45, 3480-3489; (b) J. Tan, B. Chen, L. He, Y. Tang, Z. Jiang, G. Yin, J. Wang, X. Jiang. Anacardic acid (6-pentadecylsalicylic acid) induces apoptosis of prostate cancer cells through inhibition of androgen receptor and activation of p53 signaling. *Chin. J. Cancer Res.* **2012**, 24, 275-283.